

StepOne Plus

real-time PCR system
即時定量系統

金萬林企業股份有限公司
劉儀君 Jessie

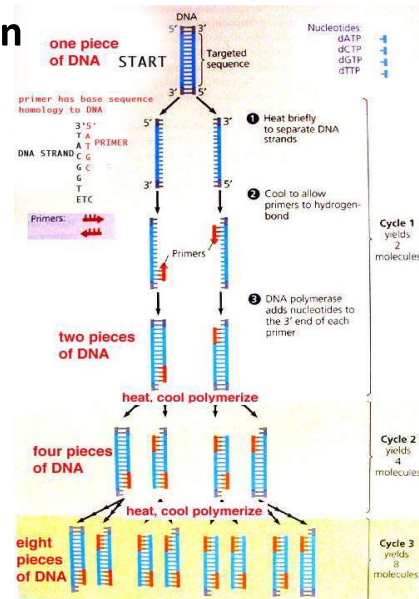
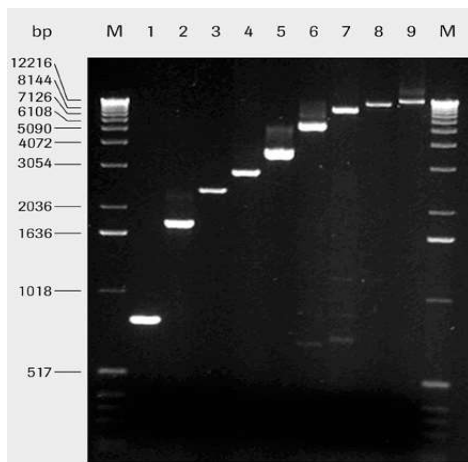
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Agenda

- Principle of real-time PCR
- Real-time PCR chemistries
- Real-time PCR Quantitation methods and applications
- Standalone operating, PC-controlled operating and data analyzed

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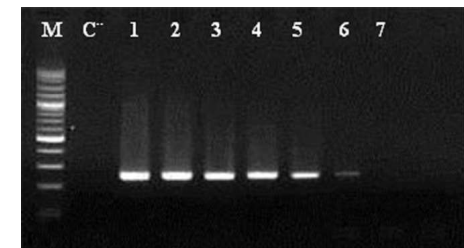
Polymerase Chain Reaction (PCR)



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Conventional PCR Quantitative System

- Low resolution
- Limited dynamic
- Semi-quantitative
- Many manual steps
- Labor intensive
- Low throughput

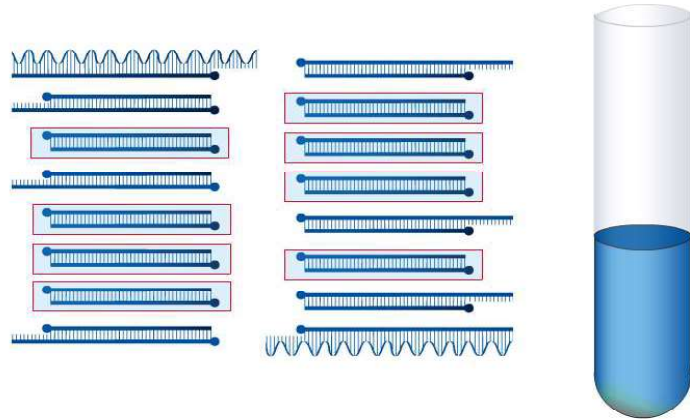


M: Marker
Lane 1: 42 pg
Lane 2: 4.2 pg
Lane 3: 420 fg
C: Control
Lane 4: 42 fg
Lane 5: 4.2 fg
Lane 6: 0.42 fg
Lane 7: 0.042 fg

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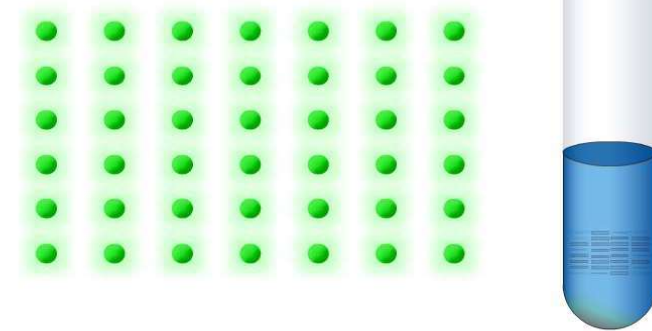
Real-Time PCR System

To perform Real-Time PCR, start with a basic PCR mix.



Real-Time PCR System

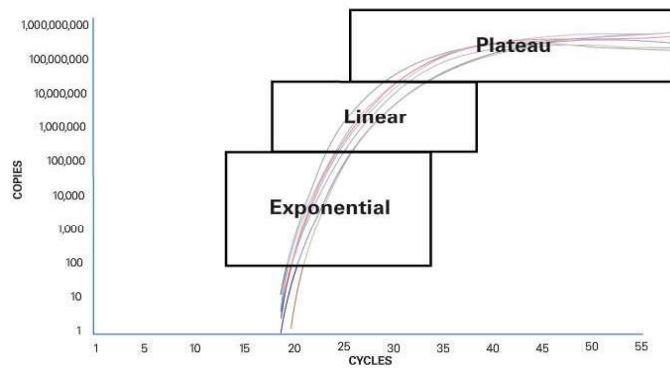
Add fluorescent labels to the PCR mix. A light source in the Real-Time PCR instrument excites the fluorescence. A camera captures the fluorescent signals.



Polymerase Chain Reaction

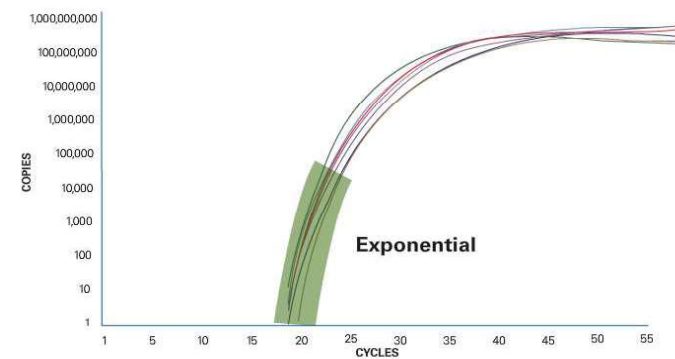
With Real-Time PCR, there are three amplification stages:

1) Exponential, 2) Linear, and 3) Plateau.



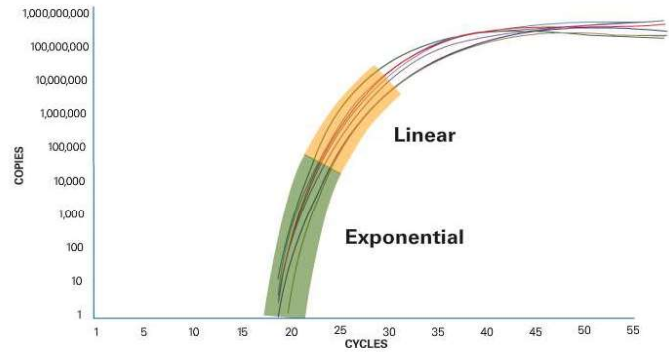
Polymerase Chain Reaction

In the Exponential phase, the reagents are in abundance and the PCR product doubles every cycle.



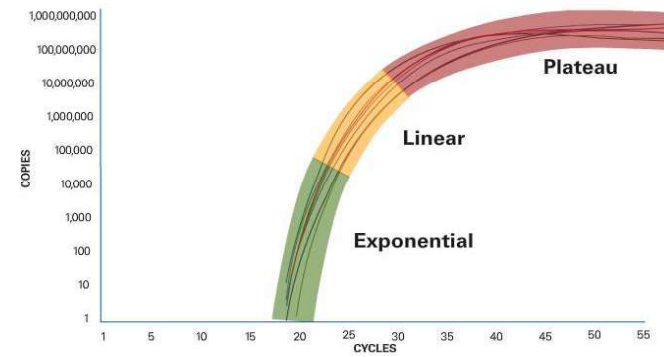
Polymerase Chain Reaction

In the Linear phase, the reagents begin to run out. The PCR reaction slows down.



Polymerase Chain Reaction

In the Plateau phase, the reagents are depleted and the PCR reaction stops.

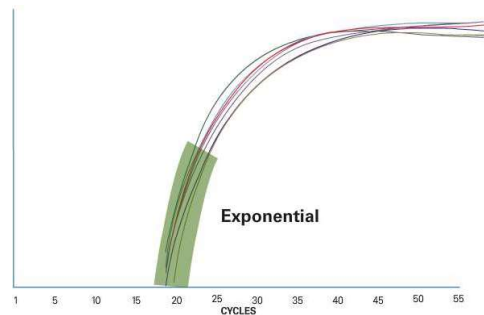


Real-Time PCR System

If PCR efficiency is 100% $\rightarrow Y = X \cdot 2^n$

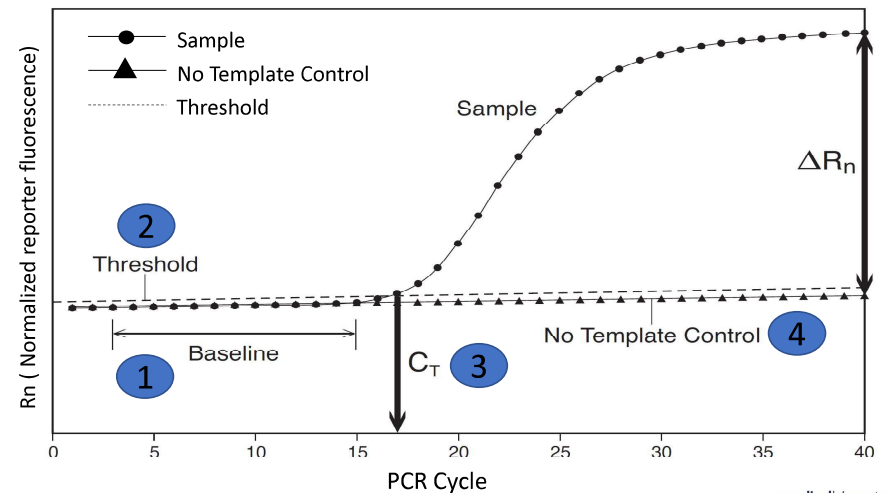
$$Y = X(1 + e)^n$$

Y: 經n個cycle後, PCR product 之總產量
X: Gene起始濃度(原始表現量)
e: Efficiency
n: PCR cycle number, Ct value

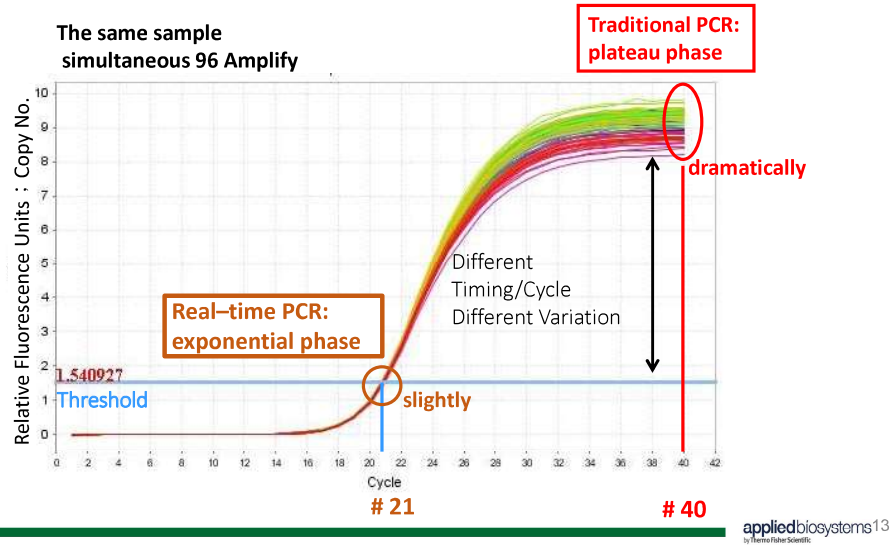


Real-Time PCR System

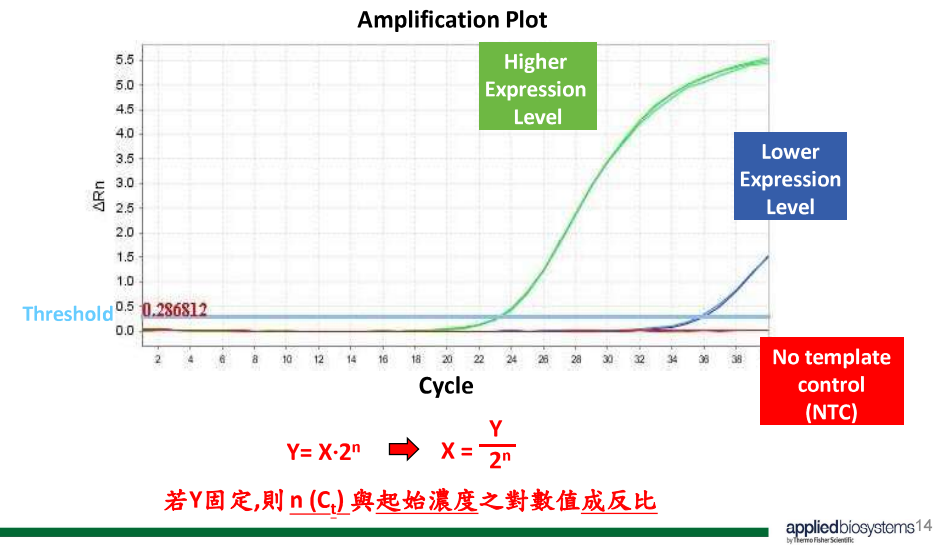
Amplification plot (linear scale)



Real-time PCR signal detection: Exponential phase



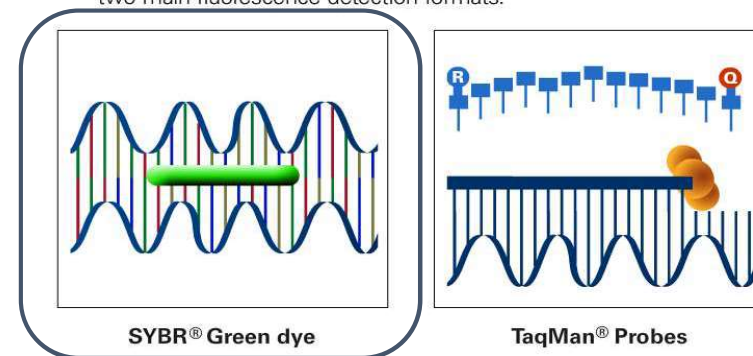
Real-Time PCR System



Real-time PCR chemistries

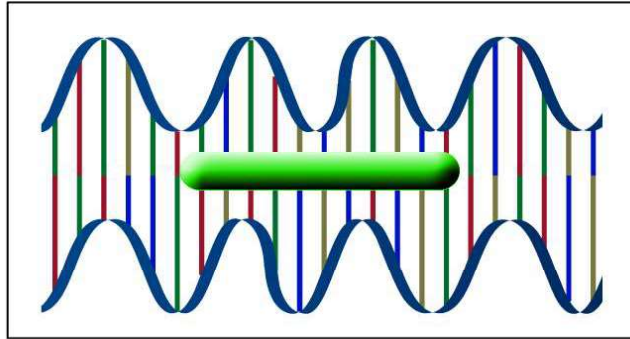
Real-Time PCR Chemistries

Real-Time PCR systems from Applied Biosystems use two main fluorescence detection formats.



Real-Time PCR System – SYBR® Green System

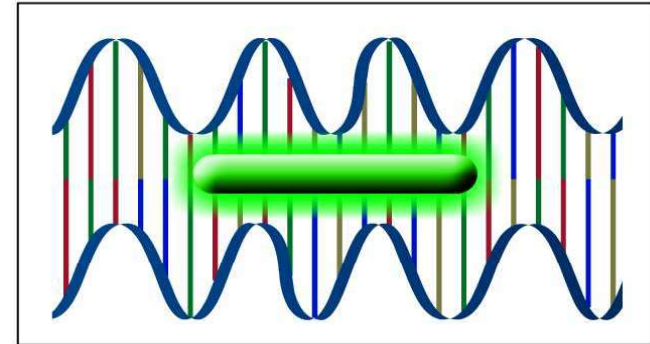
The SYBR® fluorescence format uses a dye called SYBR® Green, which binds non-specifically to double-stranded DNA.



SYBR® Green dye

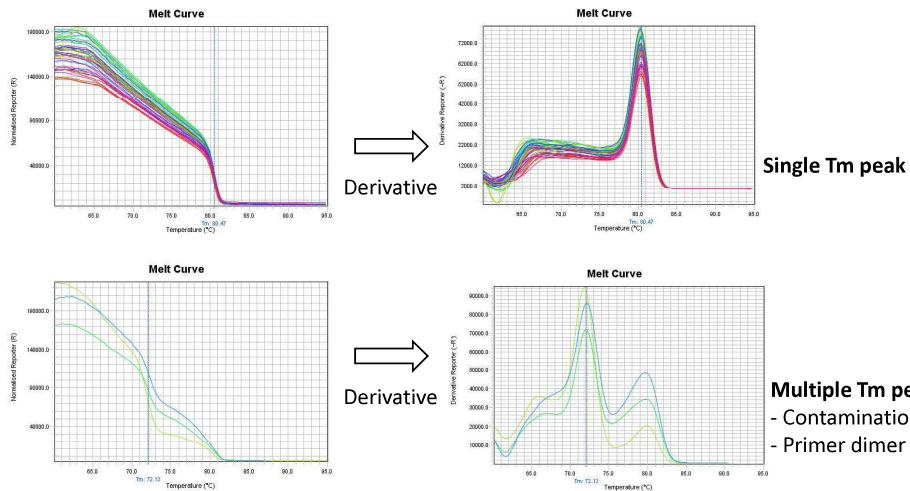
Real-Time PCR System – SYBR® Green System

The DNA-dye complex emits green light, which is recorded by the Real-Time PCR instrument.



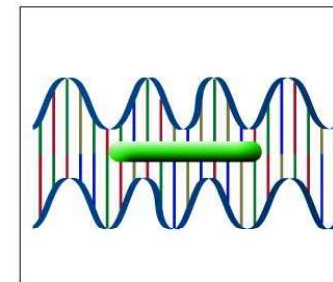
SYBR® Green dye

Real-Time PCR System – SYBR® Green System (Melting Curve)

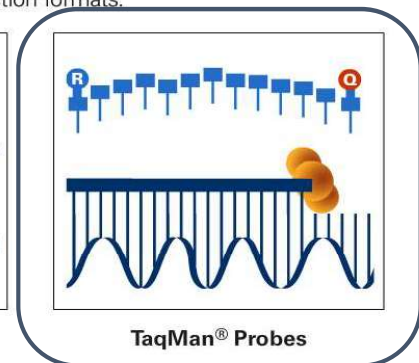


Real-Time PCR System

Real-Time PCR systems from Applied Biosystems use two main fluorescence detection formats.



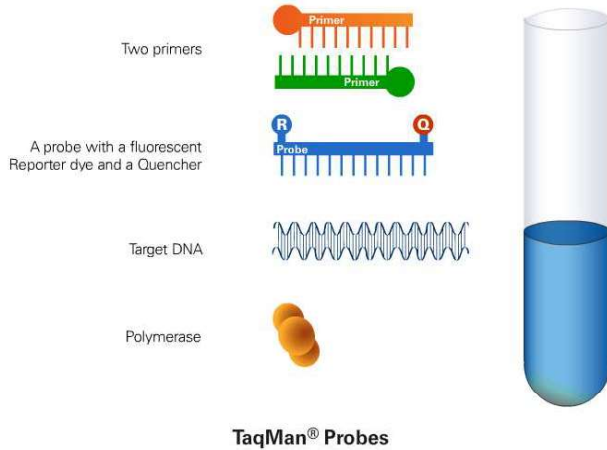
SYBR® Green dye



TaqMan® Probes

Real-Time PCR System – TaqMan® Probe System

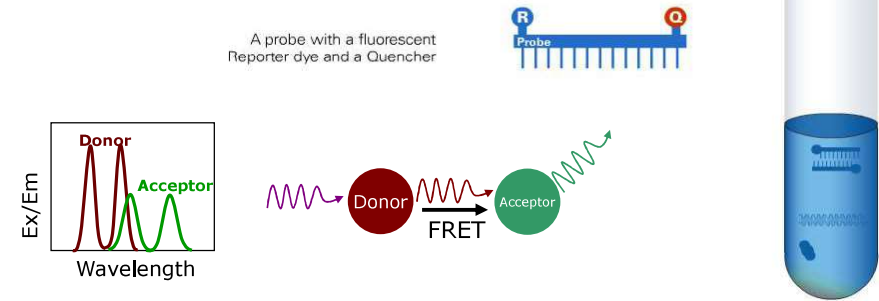
The TaqMan® Probe fluorescence format uses:



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Real-Time PCR System – TaqMan® Probe System

The design of the probe is key. The TaqMan® probe is an oligonucleotide that contains a fluorescent reporter dye bound to the 5' end and a quencher on the 3' end.

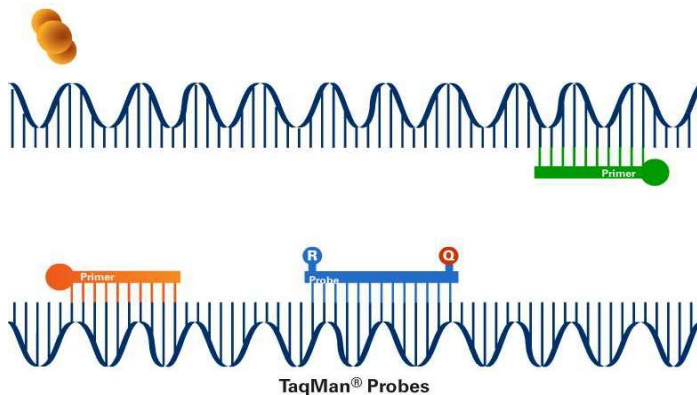


Fluorescence Resonance Energy Transfer (FRET)

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Real-Time PCR System – TaqMan® Probe System

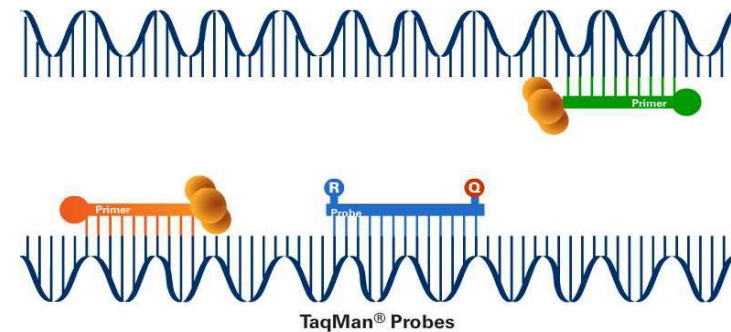
While the dye and quencher are intact, there is no fluorescence.



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Real-Time PCR System – TaqMan® Probe System

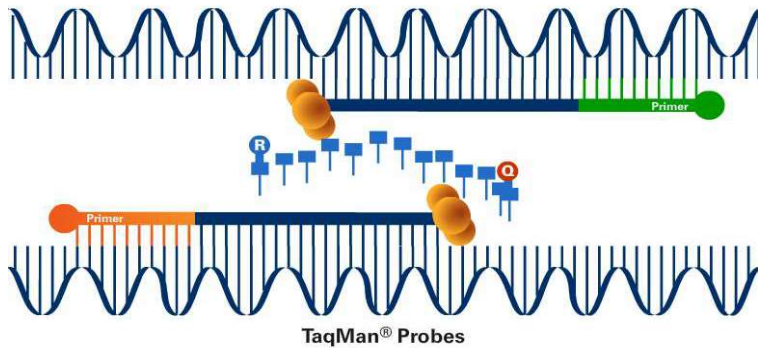
While the dye and quencher are intact, there is no fluorescence.



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Real-Time PCR System – TaqMan® Probe System

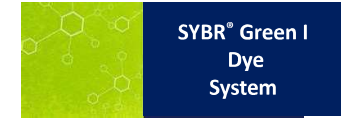
When the polymerase elongates, it is able to cleave the probe, separate the reporter from its quencher, and fluoresce. This fluorescent signal is captured by Real-Time PCR.



Real-Time PCR System (SYBR® Green I vs. TaqMan probe system)

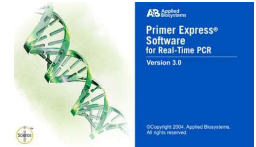


TaqMan®
Probe
System



SYBR® Green I
Dye
System

- | | | |
|---------------------|---|---|
| Specificity | <ul style="list-style-type: none"> • Highly specific • Probe Hybridization | Less specific |
| Sensitivity | <ul style="list-style-type: none"> • Very High | Very High |
| Flexibility | <ul style="list-style-type: none"> • Multiplex PCR • SNP detection • +/- application | <ul style="list-style-type: none"> • No Probe is required • Screening tool |
| Optimization | <ul style="list-style-type: none"> • Universal Guideline • Optimized 20x probe/primer mix • PCR efficiency 100±10% | <ul style="list-style-type: none"> • Universal Guideline • Need to optimize PCR program、PCR efficiency • Need to check primer-dimer information (Melting curve) |

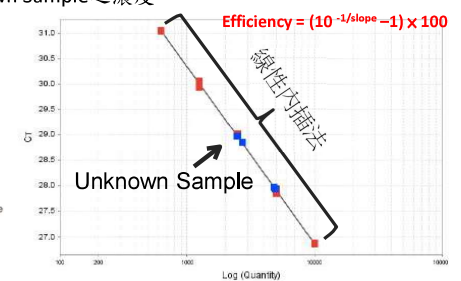
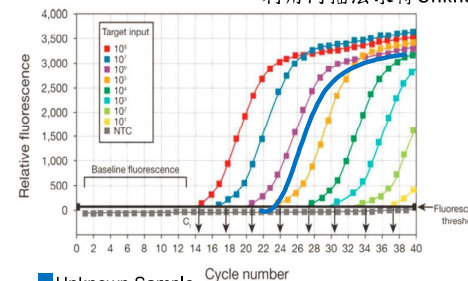


Real-time PCR Quantitation methods and applications

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Absolute Quantitation

已知濃度Standard Sample繪製標準曲線
利用內插法求得Unknown Sample之濃度

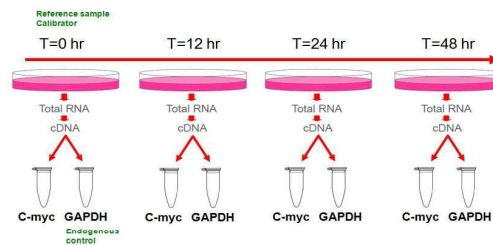


Unknown Sample	Unknown Sample
Target: RNase P Slope: -3.477 Y-inter: 40.768 R ² : 0.999 Eff%: 93.912	Target: RNase P Slope: -3.477 Y-inter: 40.768 R ² : 0.999 Eff%: 93.912
Standard	Unknown
Unknown (Flagged)	Unknown (Flagged)

Problem	Possible cause and solution
PCR efficiency is >110%	Non-specific products. Use melting curve analysis and gel electrophoresis to identify non specific amplicons. Optimize your primer design to avoid such artifacts or use validated pre-designed primers.
PCR efficiency is <90%	PCR inhibitors present in a reaction mixture. Re-purify your template DNA. PCR conditions are suboptimal. Primer design. Verify the primer concentrations. Verify storage conditions of qPCR master mix. Verify your primer design, use primer design programs or validated pre-designed primers. Avoid designing primers in regions with high DNA secondary structure.

- Standard Sample
- When slope = -3.32 Efficiency = 100%
 - The optimal efficiency range: 90-110%
 - R-value should be ≥0.99

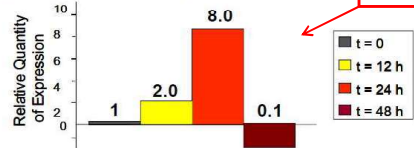
Relative Quantitation (comparative Ct)



- ✓ Target gene: *c-myc*
- ✓ Reference gene (Endogenous control): *GAPDH*
- ✓ Calibrator: T=0 hr

$\Delta\Delta C_t$ Calculations (Comparative C_t)

	c-Myc	GAPDH	ΔC_t	$\Delta\Delta C_t$	$2^{-\Delta\Delta C_t}$
T=0 (calibrator)	25	10	15	0	1.0
T=12hr	24	10	14	-1	2.0
T=24hr	23	11	12	-3	8.0
T=48hr	28	10	18	3	0.1



step 1: Normalization to endogenous control

Sample: $Ct\ c-myc - Ct\ GAPDH = \Delta C_t\ sample$
 Calibrator: $Ct\ c-myc - Ct\ GAPDH = \Delta C_t\ calibrator$

step 2: Normalization to calibrator sample

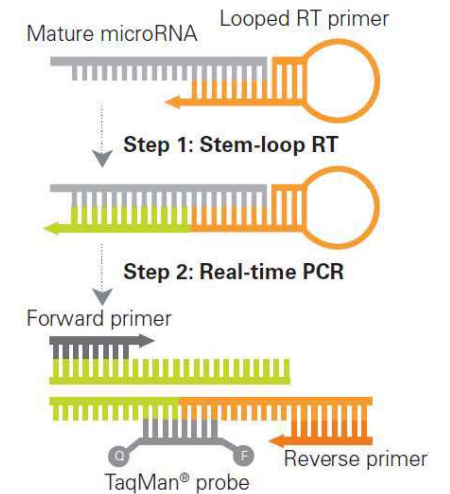
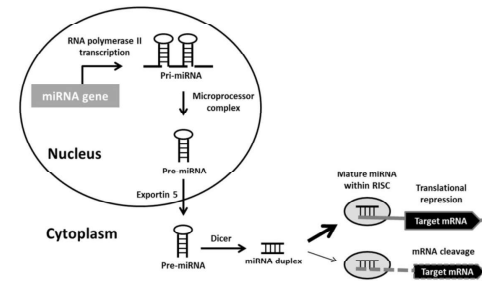
$\Delta C_t\ Sample - \Delta C_t\ Calibrator = \Delta\Delta C_t$

step 3: use the formula

$$2^{-\Delta\Delta C_t}$$

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TaqMan microRNA Assays



- Each assay contain:
 - miRNA-specific RT primer
 - miRNA-specific TaqMan Assay
- human, mouse, rat, *Drosophila*, *C. elegans*, and *Arabidopsis* etc. available; coverage for miRBase v.21

Front Cell Neurosci, Oct 2013 vol.7, article178
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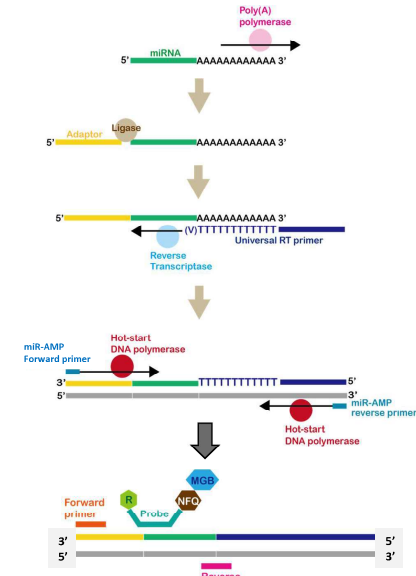
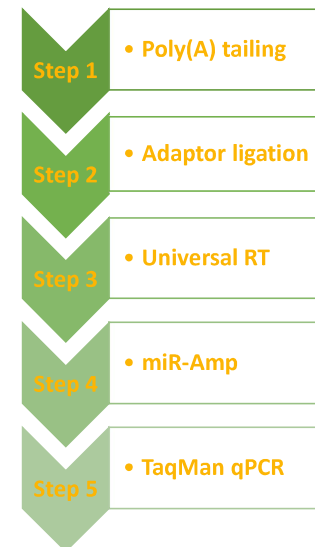
TaqMan Advanced miRNA Assays

- superior sensitivity in tissue, serum, and plasma
- 1 ng of total RNA from tissue or 2 μ L of eluant from serum or plasma
- Have multiple miRNA targets from a single amplified sample
- TaqMan Advanced miRNA Assays is compatible with TaqMan advances miRNA cDNA synthesis kit



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TaqMan Advanced miRNA Assays: workflow



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TaqMan Advanced miRNA Assays (High specificity and sensitivity)

TaqMan Advanced miRNA Assay	Synthetic template							
	Let-7a	Let-7b	Let-7c	Let-7d	Let-7e	Let-7f	Let-7g	Let-7i
Let-7a	100%	0%	0%	0%	4%	2%	0%	0%
Let-7b	0%	100%	3%	0%	0%	0%	0%	0%
Let-7c	1%	2%	100%	0%	0%	0%	0%	0%
Let-7d	0%	0%	0%	100%	0%	0%	0%	0%
Let-7e	0%	0%	0%	0%	100%	0%	0%	0%
Let-7f	1%	0%	0%	0%	0%	100%	0%	0%
Let-7g	0%	0%	0%	0%	0%	0%	100%	4%
Let-7i	0%	1%	0%	0%	0%	0%	0%	100%

miRNA name	miRNA sequence
hsa-let-7a-5p	UGA GGU AGU AGG UUG UAU AGU U
hsa-let-7b-5p	UGA GGU AGU AGG UUG UGU GGU U
hsa-let-7c-5p	UGA GGU AGU AGG UUG UAU GGU U
hsa-let-7d-5p	AGA GGU AGU AGG UUG CAU AGU U
hsa-let-7e-5p	UGA GGU AGG AGG UUG UAU AGU U
hsa-let-7f-5p	UGA GGU AGU AGA UUG UAU AGU U
hsa-let-7g-5p	UGA GGU AGU AGU UUG UAC AGU U
hsa-let-7i-5p	UGA GGU AGU AGU UUG UGC UGU U

Highly homologous members of the let-7 miRNA family

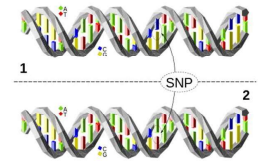
There is minimal or no cross-reactivity.

* Represent Differences in nucleotide

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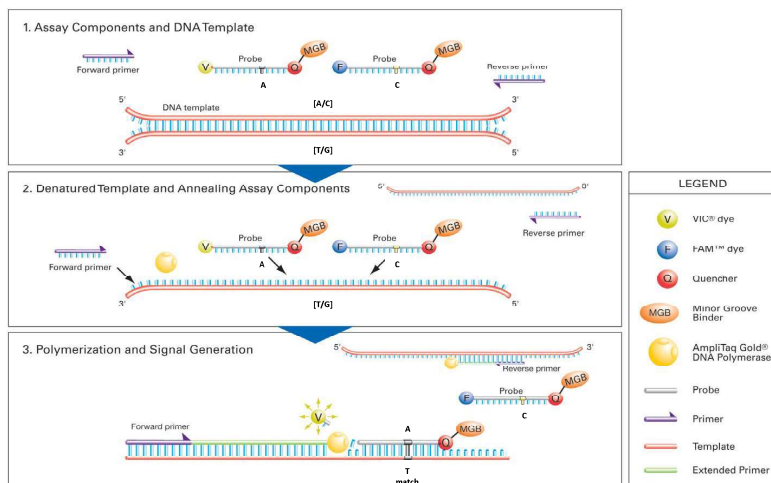
What are Single Nucleotide Polymorphism (SNP)?

- Diploid organisms – 2 sets of chromosomes
- Each person receives 1 alleles from each parent.
- If both alleles are the same, the person is **homozygous** for that gene.
- If the alleles differ, the person is **heterozygous** for that gene.
- They occur once in every 300 nucleotides on average.
- The rarer allele having a frequency of at least 1%.



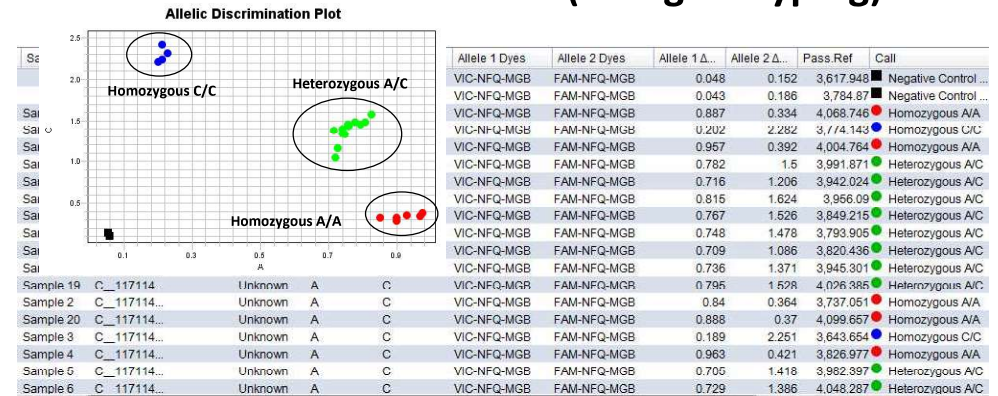
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TaqMan Probe System (Single Nucleotide Polymorphism)








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Allelic Discrimination Plot (SNP genotyping)



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Application of TaqMan Assays and SYBR systems

	TaqMan assays	SYBR Green
	<ul style="list-style-type: none"> ➢ Gene expression analysis (ex: Genetically Modified Organism, GMO) 	<ul style="list-style-type: none"> ➢ Gene expression analysis
	<ul style="list-style-type: none"> ➢ MicroRNA and noncoding RNA analysis 	
	<ul style="list-style-type: none"> ➢ Drug metabolism genotyping ➢ SNP genotyping ➢ Somatic mutation detection 	
	<ul style="list-style-type: none"> ➢ Pathogen Presence/ Absence 	
	<ul style="list-style-type: none"> ➢ Protein expression 	

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The StepOnePlus Real-Time PCR System



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Design and Feature



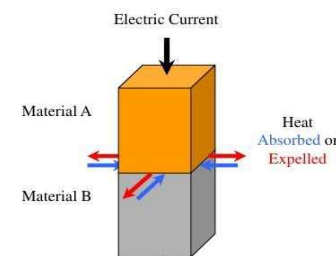
StepOnePlus 96 well

1. PCR +Optical sys. 避免光徑偏移
2. Flash LED 偵測才開啟，耗損少
3. Peltier effect 精確穩定控溫系統
4. USB 存出 (主機會暫存最後一筆檔案)



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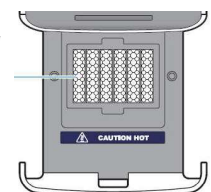
Block



Tech.	→ Ramping ←	Volume
1. Peltier effect 不同導體 電流改變 熱能吸放 熱電效應 精確控溫	1. 鎳鉻合金基座 4.6 °C/sec 2. 樣品升降溫 Fast : ± 2.2 °C/sec Std: ± 1.6 °C/sec 3. Auto delta Time by cycle Temp by cycle	1. 反應管 (0.1 mL) 96 well plate 8-tube strip single tube *耗材需使用"Optical" 2. 反應體積 10-30 µL



CAUTION



StepOnePlus instrument VeriFlex™ Sample Blocks

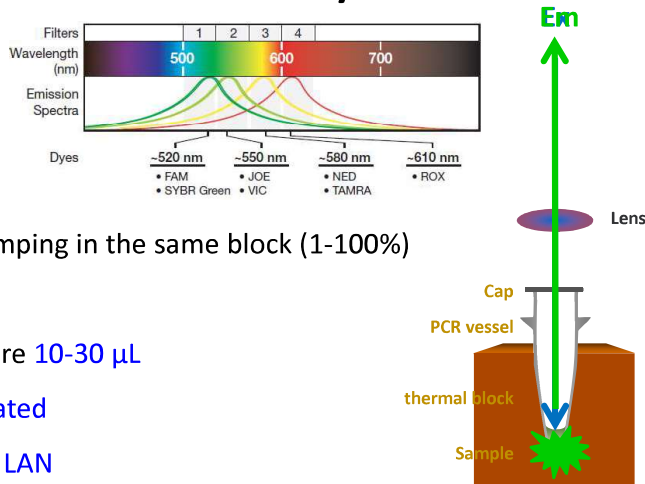
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StepOne™ Plus Real-Time PCR System

- 96-well
- 4 color system (TaqMan MGB probe)

Common Features

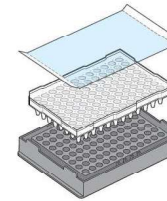
- Fast and Standard ramping in the same block (1-100%)
- Single LED excitation
- Supported volumes are 10-30 µL
- Standalone or co-located
- Remote monitoring - LAN



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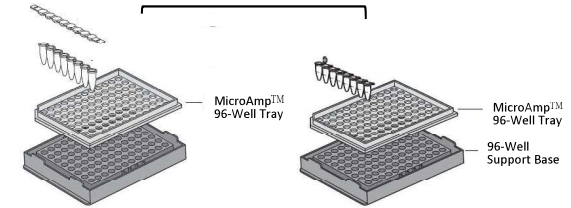
Supported Consumables

- for 0.1 mL Optical tube strip/single tube
- ✓管子及蓋子上請勿標記任何記號
避免墨色脫落沾到 PCR block增加背景值



more than 48 reactions/run

- ✓96-well plate +
Optical adhesive film



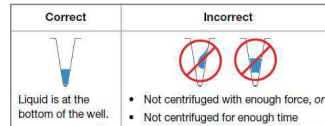
less than 48 reactions/run

- ✓使用八連排或單管必須將其置放在
MicroAmp™ 96-Well Tray
- ✓對稱放置，避免加熱板上升時，受
力不均，造成機械性的損傷。

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上機前/時之注意事項

- ✓光學封膜/瓶蓋是否平整緊密貼附，且表面無指紋或其他
髒汙沾黏。
- ✓確認已將 reaction mixture 完全離心至反應管底部，且無
氣泡產生 (Centrifuge the tubes for 2 min at less than 1500
rpm)
- ✓上機時，反應管放置位置應以
“平行、偶數、對稱”為原則。
- ✓為了避免卡盤，不建議8連排剪裁後上機使用。



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SetUp

1. PC free



2. PC controlled



or

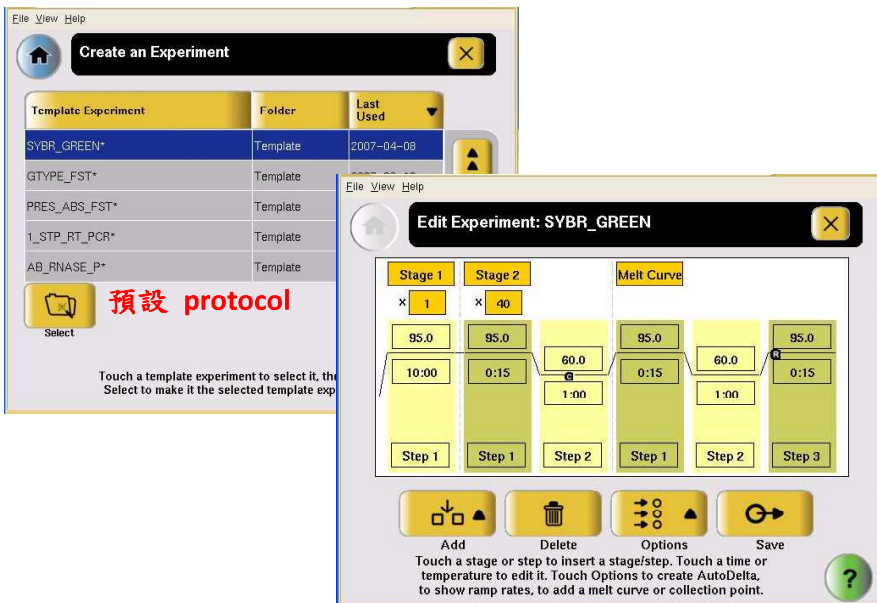
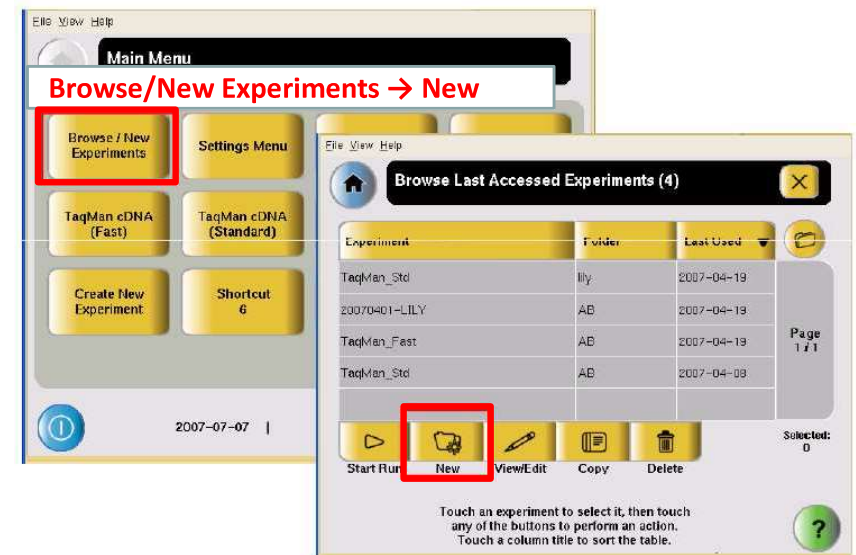


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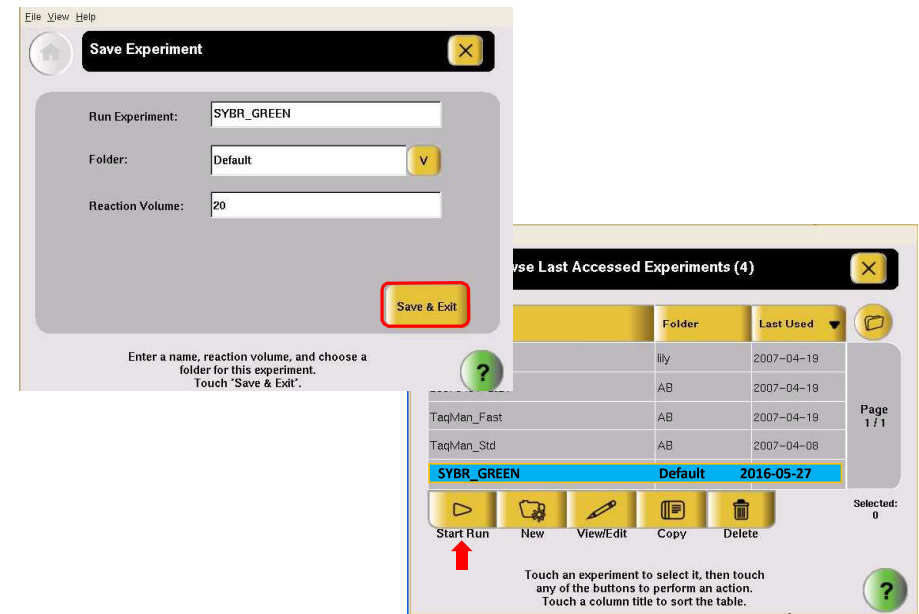
Standalone (PC-free)

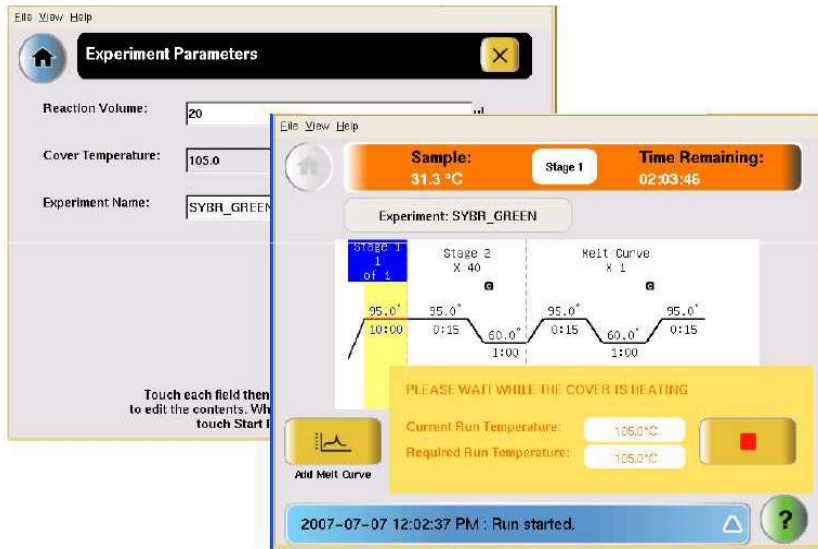


1. **Start the run** from the touch screen.
2. **Download the file (.eds)** to your PC. (主機暫存最後一筆檔案)
3. **Analyze** your data.



Select Experiment → Save



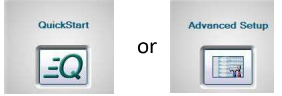


SetUp

1. PC free



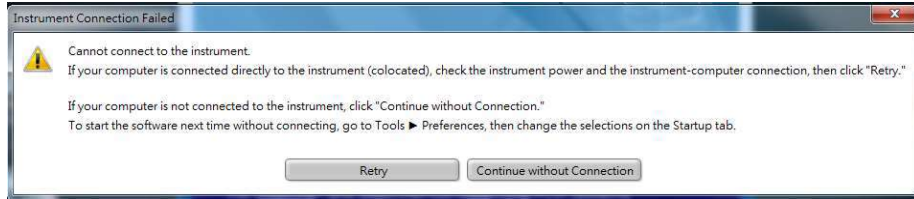
2. PC controlled



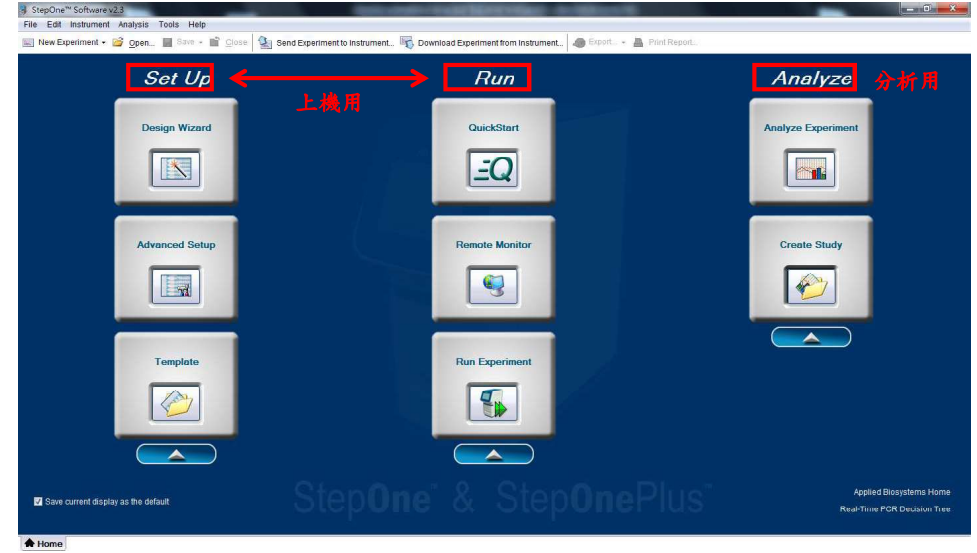
StepOne Software v2.3 軟體操作



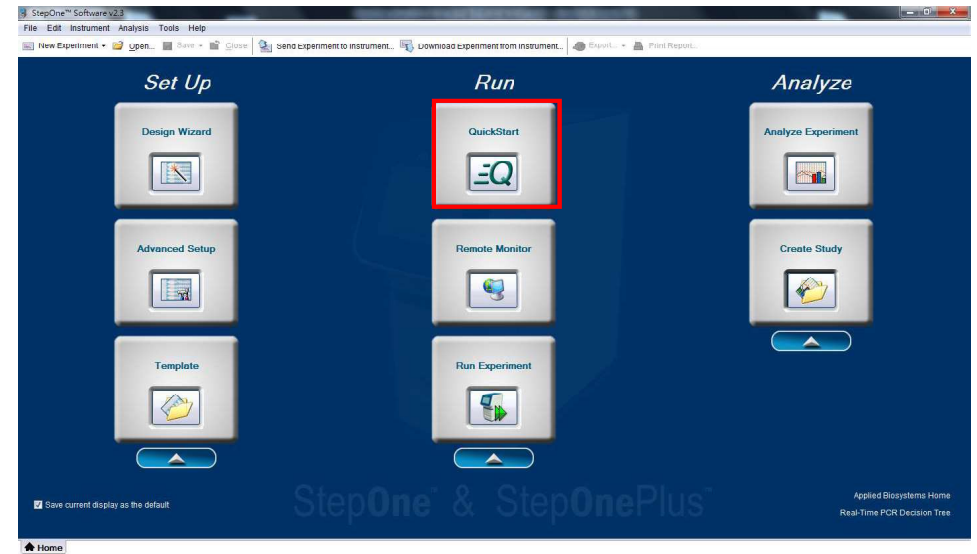
Login the software



* Analyze data -> Continue without Connection



Run the experiment



Run (QuickStart) : Experiment Properties

1. Enter Experiment Name and Location 輸入檔案名稱、選擇目錄路徑

2. Select Experiment Type 選擇實驗類型 (定性、定量)

3. Select Reagents 選擇試劑類型 (螢光系統)

4. Select Ramp Speed 選擇Ramp Speed

5. Select Template 選擇上樣板類型

Quantitative

Qualitative

START RUN

appliedbiosystems57

Run (QuickStart) : Run Method

Run Method 輸入反應體積、
確認溫度 時間 收光時間點

Reaction Volume Per Well: 20 μ L

Number of Cycles: 40

Starting Cycle: 1

95.0 °C 00:20 100%

95.0 °C 00:01 100%

90.0 °C 00:20 100%

START RUN

appliedbiosystems58

Analyze the data

Analyze: Analyze Experiment

Set Up Run Analyze

Design Wizard QuickStart Analyze Experiment

Advanced Setup Remote Monitor Create Study

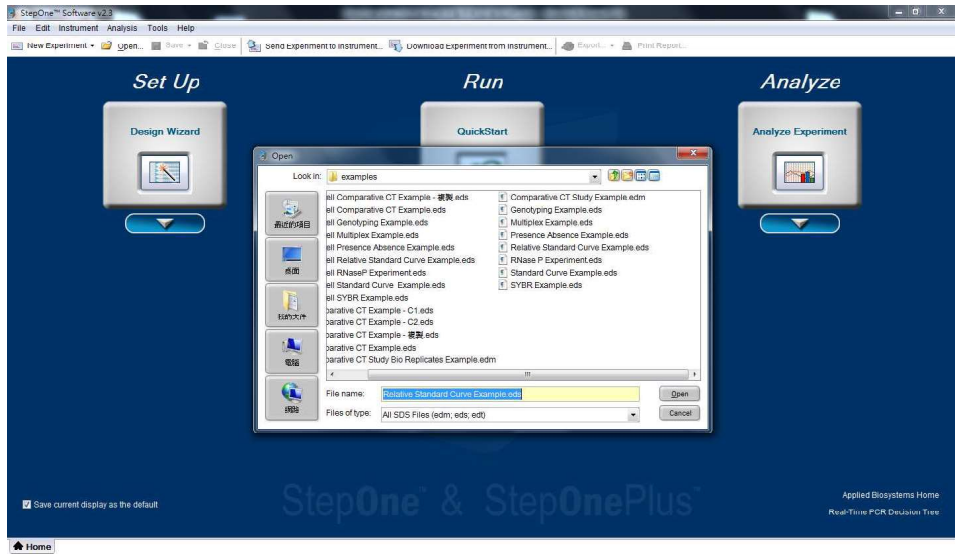
Template Run Experiment

START RUN

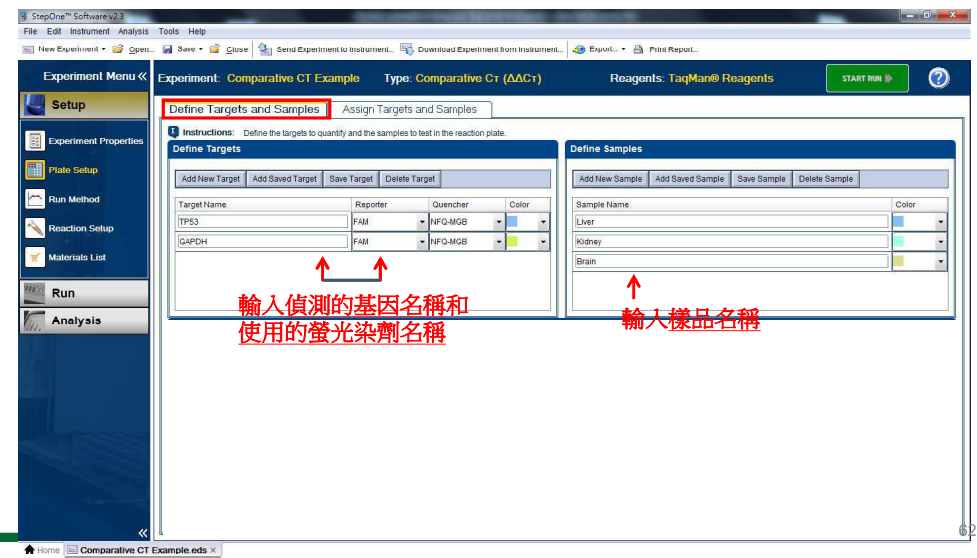
StepOne & StepOnePlus

Applied Biosystems Home
Real-Time PCR Decision Tree

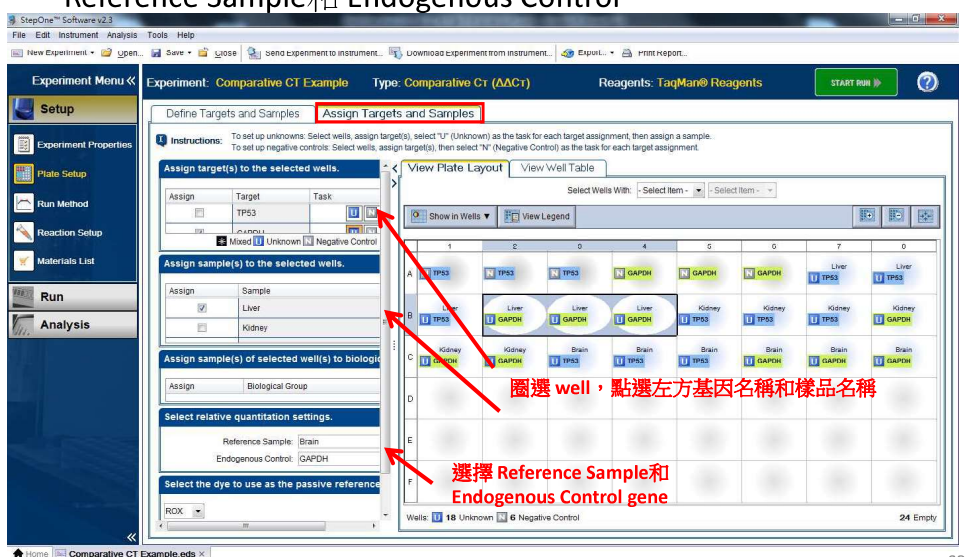
appliedbiosystems60



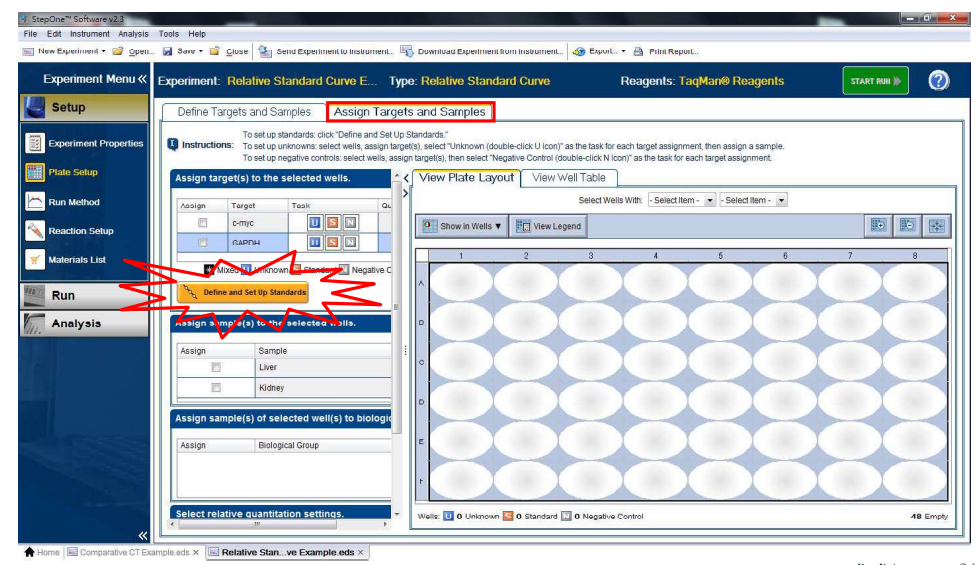
Setup: plate Setup → Define Targets and Samples 定義偵測的基因和樣品名稱



Setup: plate Setup → Assign Targets and Samples 定義基因和樣品位置、設定 Reference Sample 和 Endogenous Control



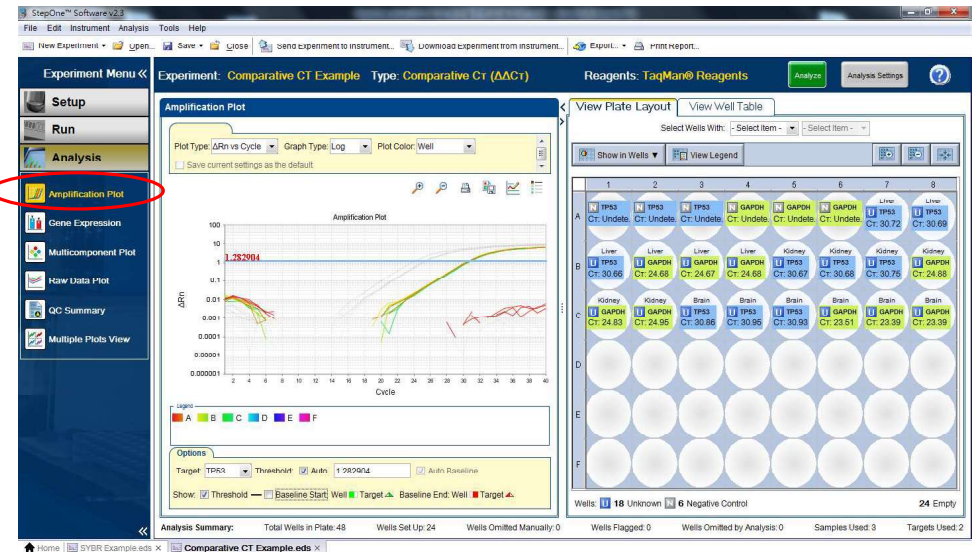
Setup: plate Setup → Assign Targets and Samples 定義標準品樣品位置、濃度



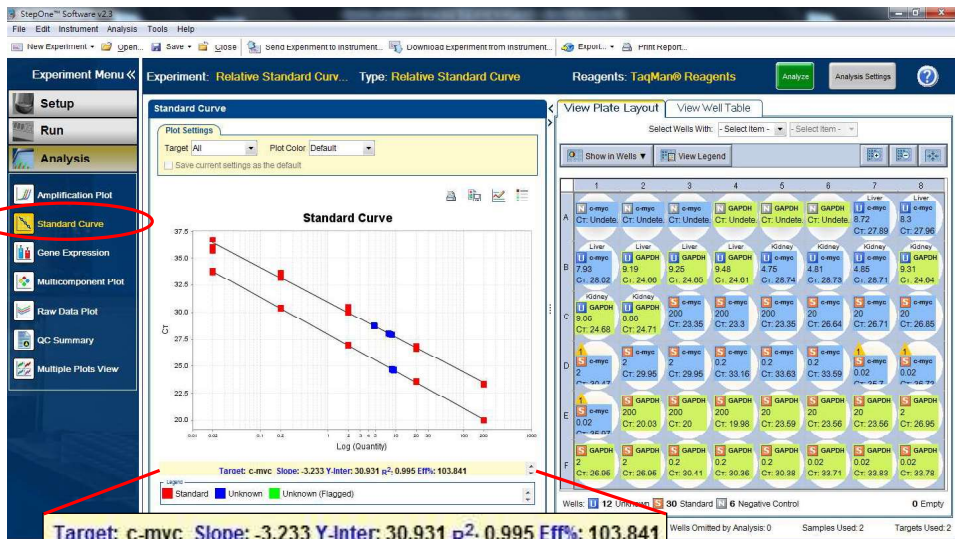
Analysis: Melt curve (SYBR Green only)



Analysis: Amplification plot

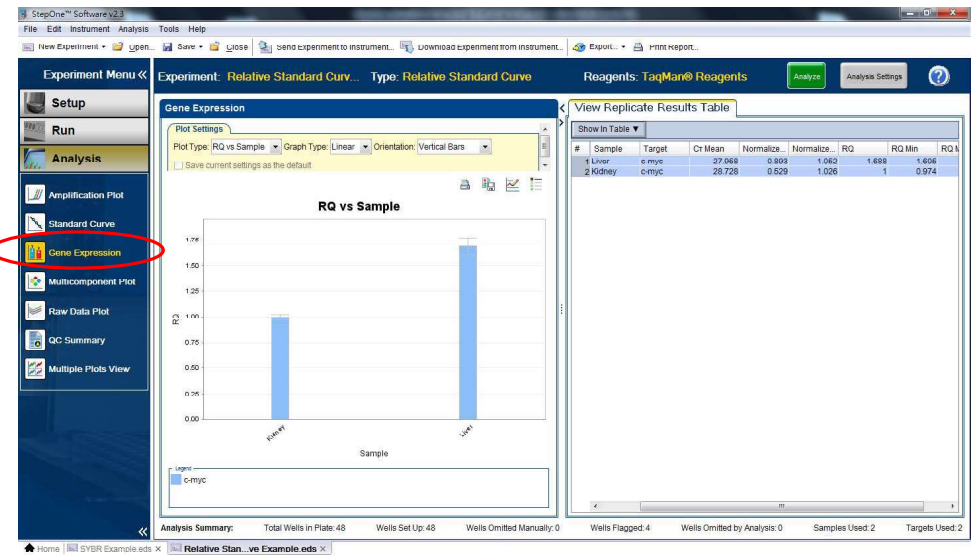


Analysis: Standard curve



確認各基因之 Efficiency 落在 90~110%

Analysis: Gene Expression



Analysis: QC Summary → 協助 troubleshooting

StepOne™ Software v2.3

Experiment: Relative Standard Curve... Type: Relative Standard Curve

Reagents: TaqMan® Reagents

QC Summary

Flag Details

Flag	Name	Frequency	Wells
AMPNC	Amplification in negative control	0	
BADPROX	Bad passive reference signal	0	
OFFSCALE	Fluorescence is offscale	0	
HIGHSD	High standard deviation in replicate group	3	D7, D8, E1
NOAMP	No amplification	0	
RUNISE	Noise higher than others in plate	0	
SPIKE	Noise spikes	0	
NO SIGNAL	No signal in well	0	
OUTLIERRG	Outlier in replicate group	1	D1
EXPFAIL	Exponential algorithm failed	0	
BLFAIL	Baseline algorithm failed	0	
THOLDFAIL	Thresholding algorithm failed	0	

Flag: HIGHSD—High standard deviation in replicate group
Flag Detail: The Ct standard deviation for the replicate group exceeds the flag setting.
Flag Criteria: Ct standard deviation > 0.5
Flagged Wells: D7, D8, E1
[View HIGHSD Troubleshooting Information](#)

Wells: 12 Unknown 30 Standard 6 Negative Control 0 Empty

Analysis Summary: Total Wells in Plate: 48 Wells Set Up: 48 Wells Omitted Manually: 0 Wells Flagged: 4 Wells Omitted by Analysis: 0 Samples Used: 2 Targets Used: 2

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by Thermo Fisher Scientific

Analysis: Multiple Plots View → Export to Excel or save as JPEG

StepOne™ Software v2.3

Experiment: Relative Standard Curve... Type: Relative Standard Curve

Reagents: TaqMan® Reagents

Multiple Plots View

Amplification Plot - Amp vs Cycle

Multicomponent Plot

Standard Curve

Gene Expression Plot

Wells: 12 Unknown 30 Standard 6 Negative Control 0 Empty

Analysis Summary: Total Wells in Plate: 48 Wells Set Up: 48 Wells Omitted Manually: 0 Wells Flagged: 4 Wells Omitted by Analysis: 0 Samples Used: 2 Targets Used: 2

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Thank you for your attention!

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