



# StepOne Plus

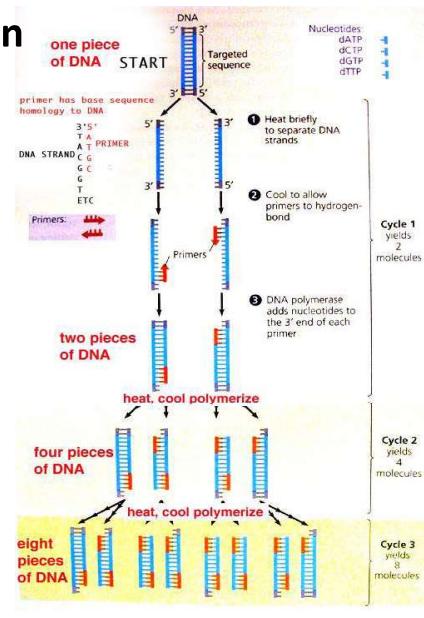
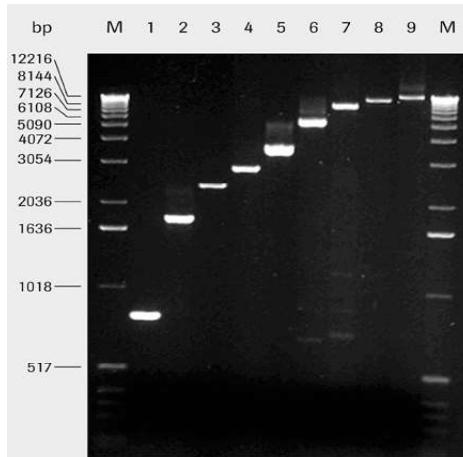
real-time PCR system  
即時定量系統

金萬林企業股份有限公司  
劉儀君 Jessie

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by Thermo Fisher Scientific

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## Polymerase Chain Reaction (PCR)



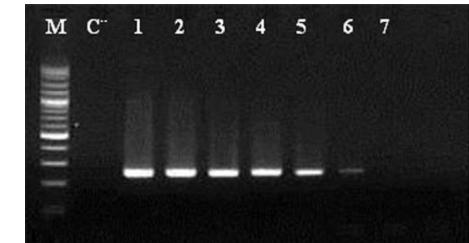
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## Agenda

- Principle of real-time PCR
- Real-time PCR chemistries
- Real-time PCR Quantitation methods and applications
- Standalone operating, PC-controlled operating and data analyzed

## Conventional PCR Quantitative System

- Low resolution
- Limited dynamic
- Semi-quantitative
- Many manual steps
- Labor intensive
- Low throughput

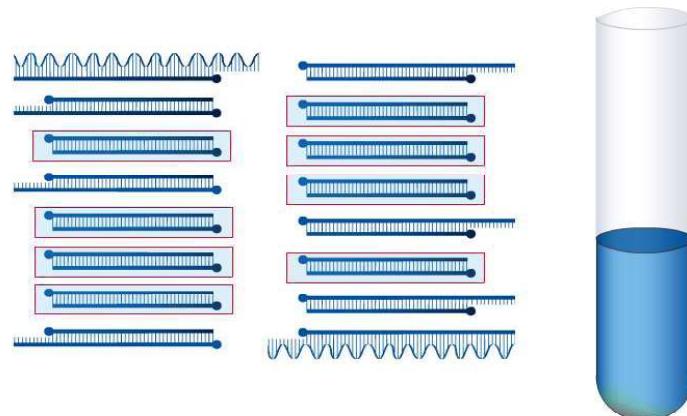


M: Marker  
C: Control  
Lane 1: 42 pg  
Lane 2: 4.2 pg  
Lane 3: 420 fg  
Lane 4: 42 fg  
Lane 5: 4.2 fg  
Lane 6: 0.42 fg  
Lane 7: 0.042 fg

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## Real-Time PCR System

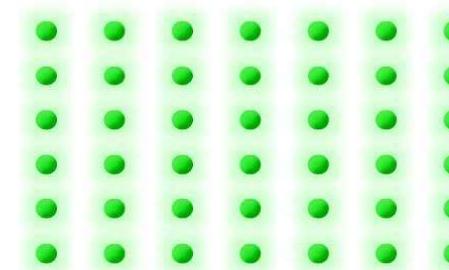
To perform Real-Time PCR, start with a basic PCR mix.



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## Real-Time PCR System

Add fluorescent labels to the PCR mix. A light source in the Real-Time PCR instrument excites the fluorescence. A camera captures the fluorescent signals.

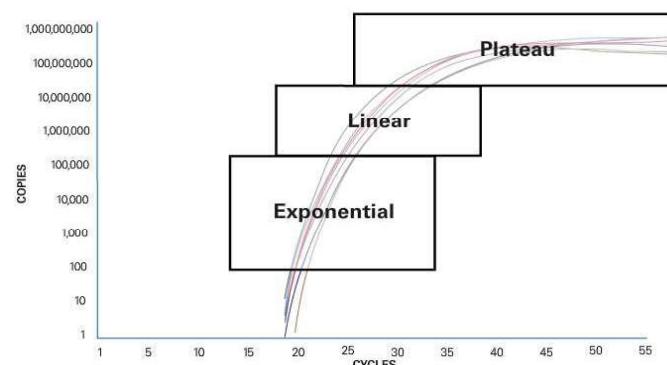


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## Polymerase Chain Reaction

With Real-Time PCR, there are three amplification stages:

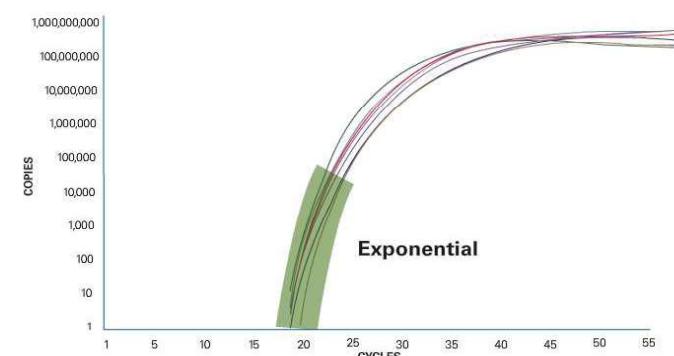
- 1) Exponential, 2) Linear, and 3) Plateau.



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## Polymerase Chain Reaction

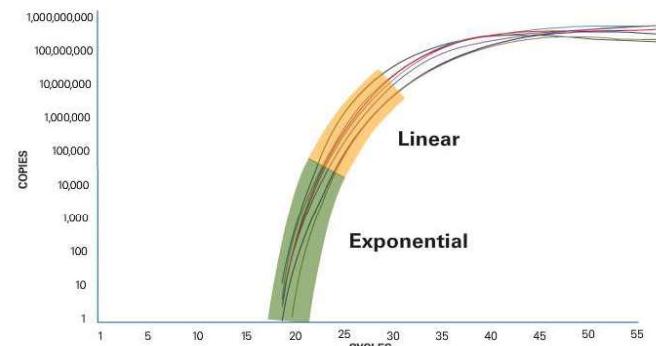
In the Exponential phase, the reagents are in abundance and the PCR product doubles every cycle.



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## Polymerase Chain Reaction

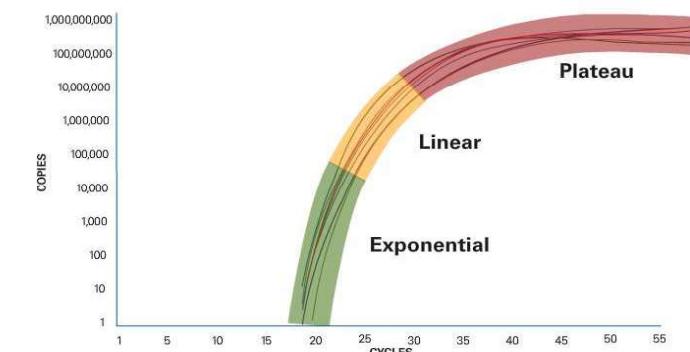
In the Linear phase, the reagents begin to run out. The PCR reaction slows down.



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## Polymerase Chain Reaction

In the Plateau phase, the reagents are depleted and the PCR reaction stops.



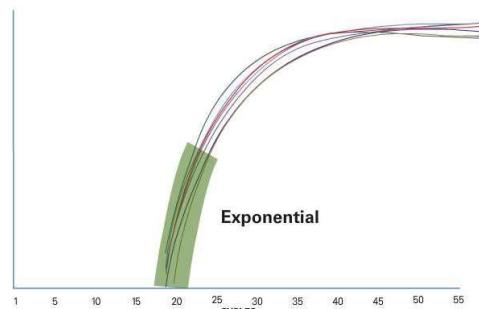
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## Real-Time PCR System

If PCR efficiency is 100%  $\rightarrow Y = X \cdot 2^n$

$$Y = X (1 + e)^n$$

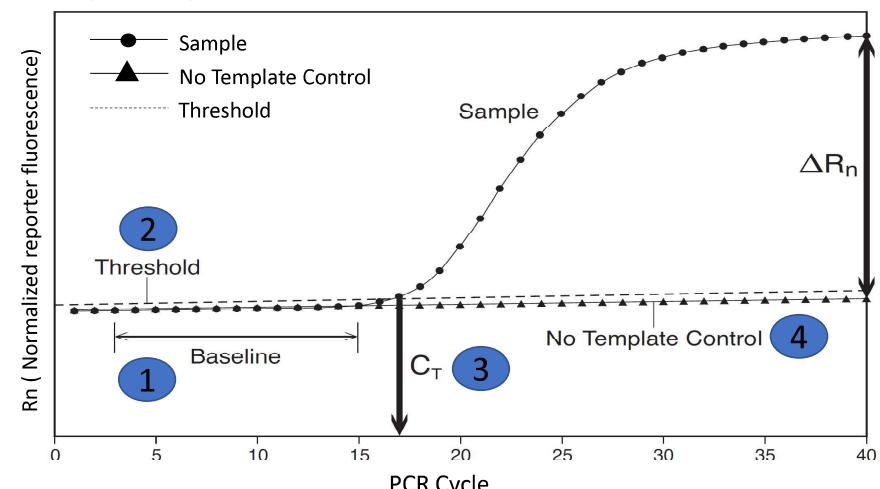
Y: 經n個cycle後, PCR product 之總產量  
X: Gene起始濃度(原始表現量)  
e: Efficiency  
n: PCR cycle number, Ct value



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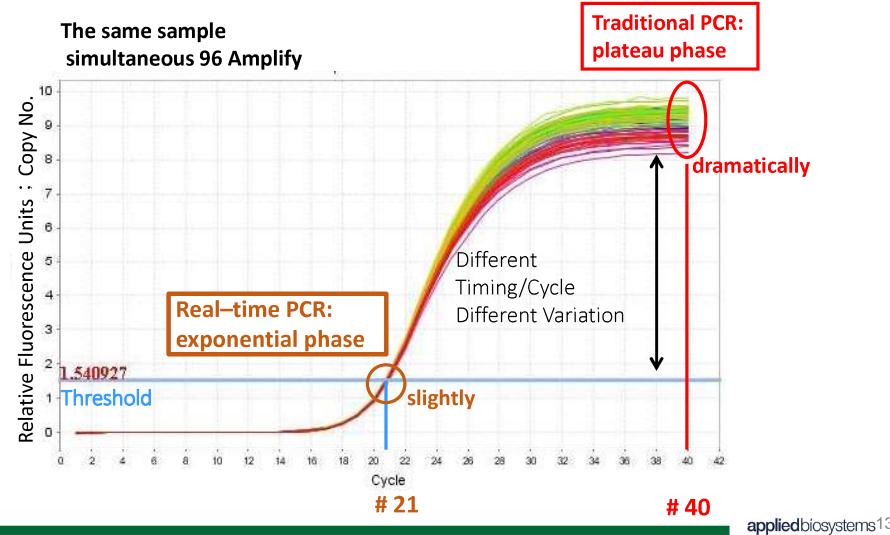
## Real-Time PCR System

Amplification plot (linear scale)

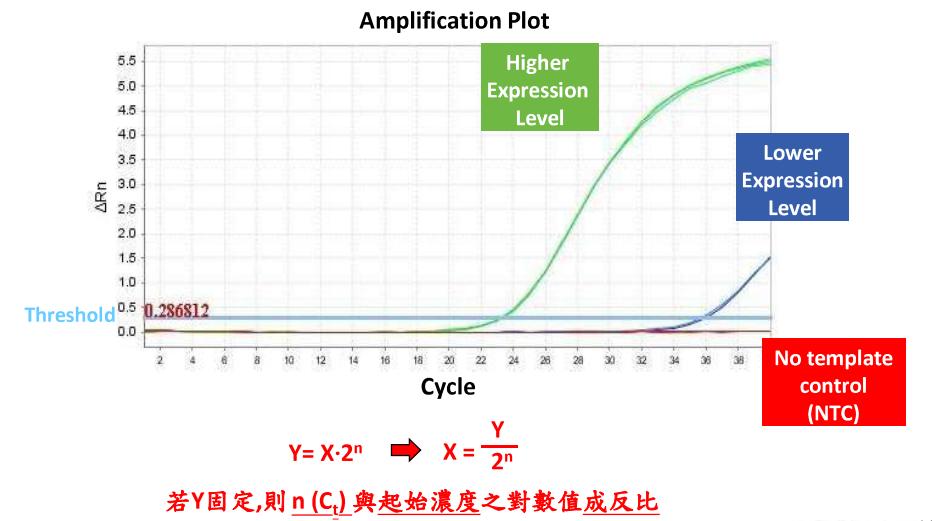


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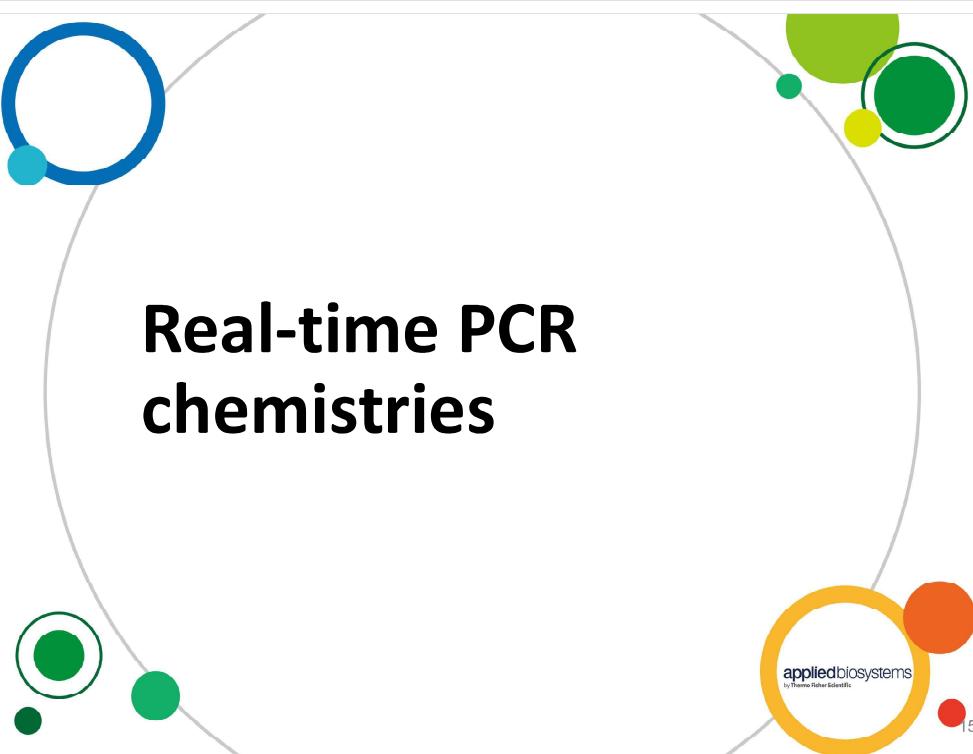
## Real-time PCR signal detection: Exponential phase



## Real-Time PCR System

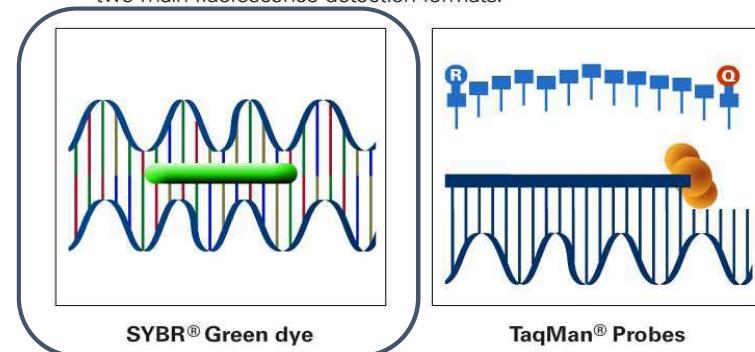


## Real-time PCR chemistries



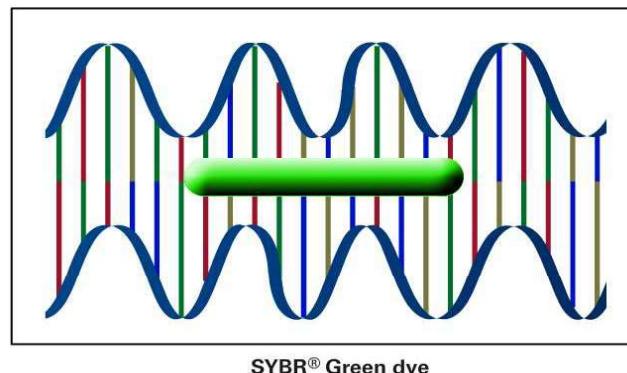
## Real-Time PCR Chemistries

Real-Time PCR systems from Applied Biosystems use two main fluorescence detection formats.



## Real-Time PCR System – SYBR® Green System

The SYBR® fluorescence format uses a dye called SYBR® Green, which binds non-specifically to double-stranded DNA.

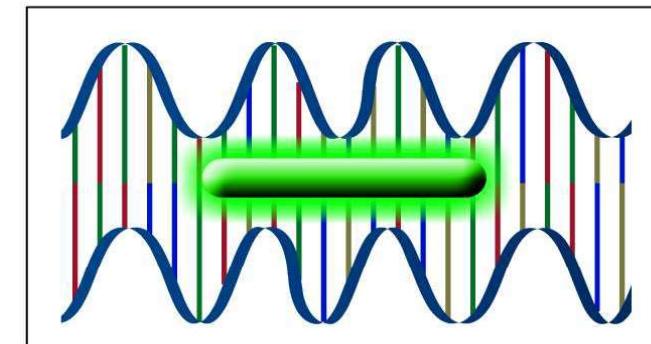


SYBR® Green dye

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## Real-Time PCR System – SYBR® Green System

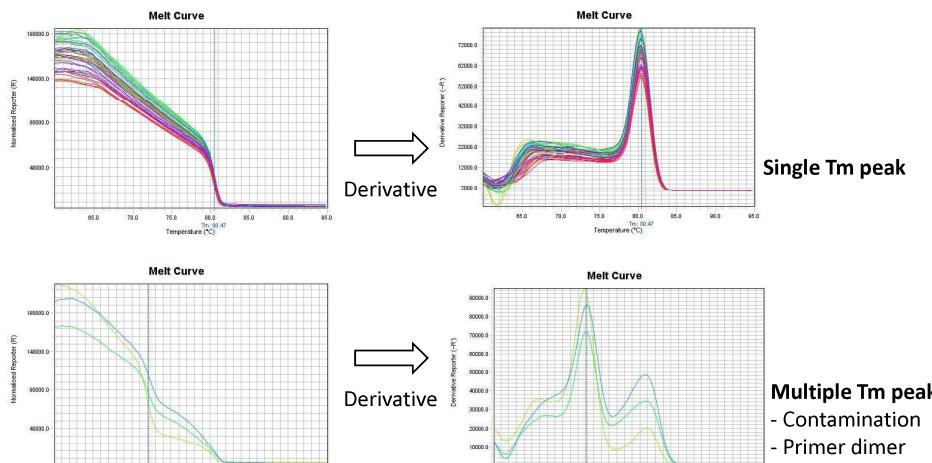
The DNA-dye complex emits green light, which is recorded by the Real-Time PCR instrument.



SYBR® Green dye

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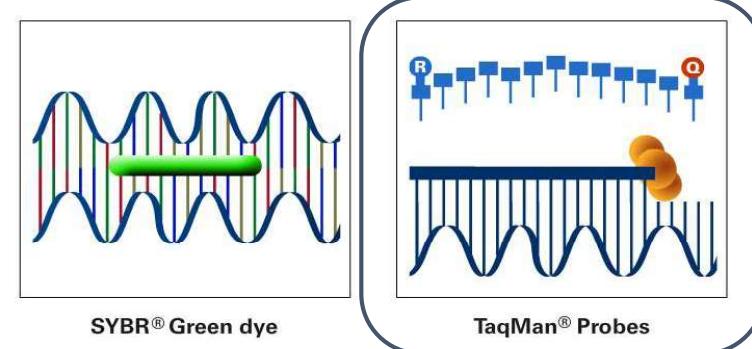
## Real-Time PCR System – SYBR® Green System (Melting Curve)



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## Real-Time PCR System

Real-Time PCR systems from Applied Biosystems use two main fluorescence detection formats.

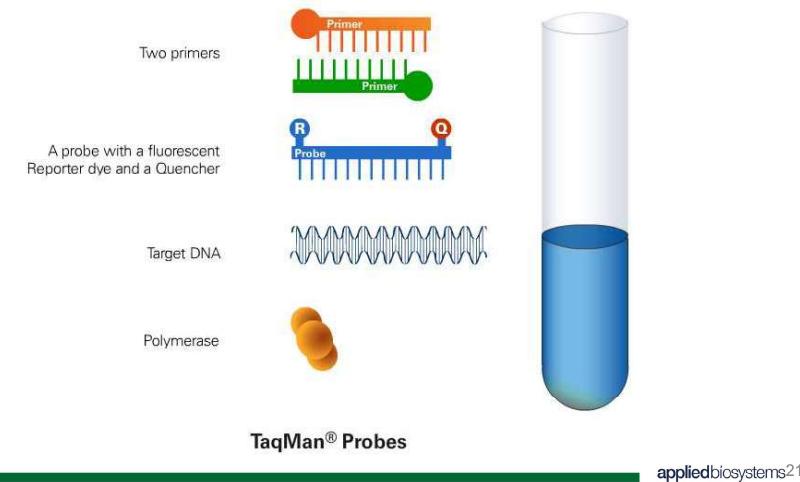


TaqMan® Probes

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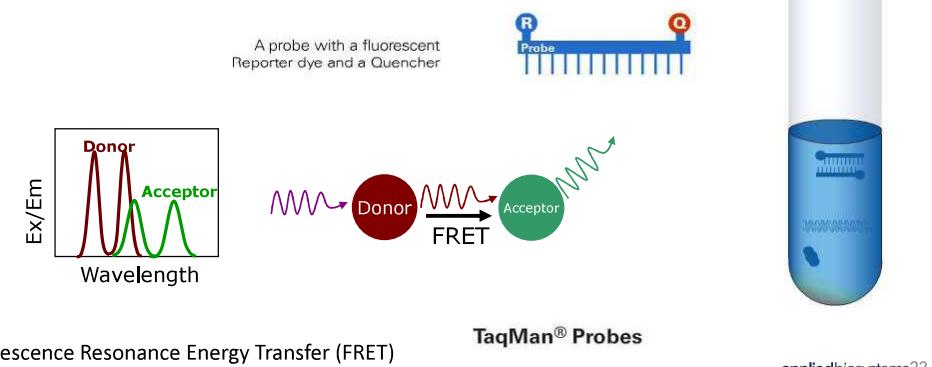
## Real-Time PCR System – TaqMan® Probe System

The TaqMan® Probe fluorescence format uses:



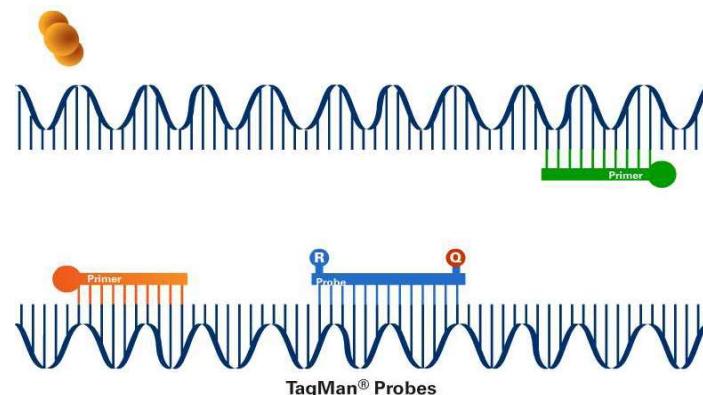
## Real-Time PCR System – TaqMan® Probe System

The design of the probe is key. The TaqMan® probe is an oligonucleotide that contains a fluorescent reporter dye bound to the 5' end and a quencher on the 3' end.



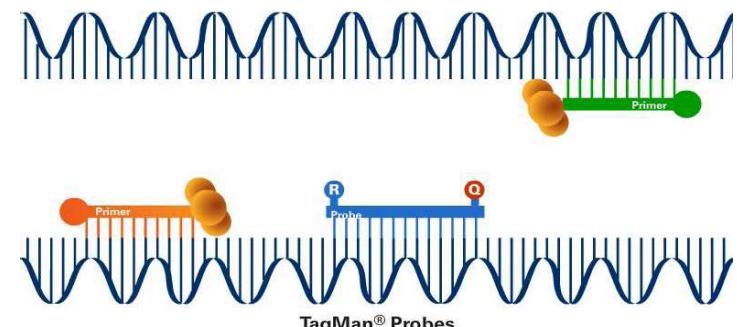
## Real-Time PCR System – TaqMan® Probe System

While the dye and quencher are intact, there is no fluorescence.



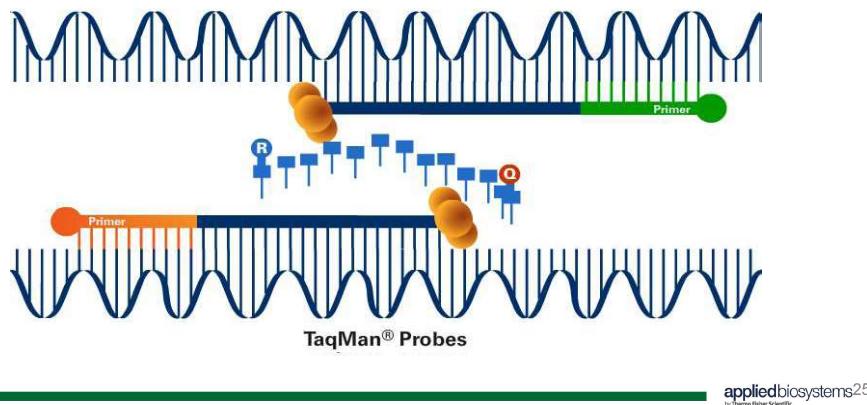
## Real-Time PCR System – TaqMan® Probe System

While the dye and quencher are intact, there is no fluorescence.



## Real-Time PCR System – TaqMan® Probe System

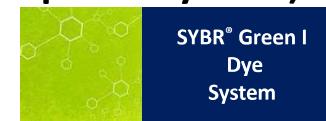
When the polymerase elongates, it is able to cleave the probe, separate the reporter from its quencher, and fluoresce. This fluorescent signal is captured by Real-Time PCR.



## Real-time PCR Quantitation methods and applications



## Real-Time PCR System (SYBR® Green I vs. TaqMan probe system)



### Specificity

- Highly specific

### Sensitivity

- Very High

### Flexibility

- Multiplex PCR
- SNP detection
- +/- application

### Optimization

- Universal Guideline
- Optimized 20x probe/primer mix
- PCR efficiency  $100 \pm 10\%$

- Less specific

- Very High

- No Probe is required
- Screening tool



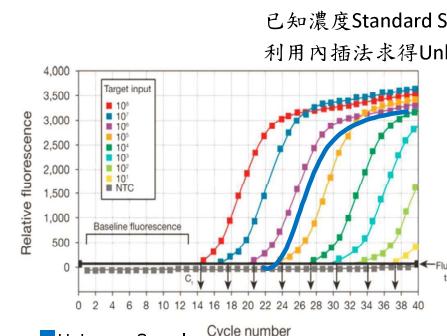
- Universal Guideline

- Need to **optimize PCR program**、**PCR efficiency**

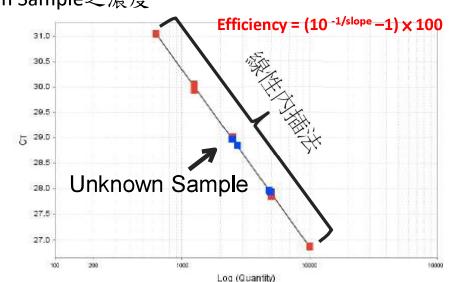
- Need to check primer-dimer information  
**(Melting curve)**

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## Absolute Quantitation



Problem	Possible cause and solution
PCR efficiency is >110%	Non-specific products. Use melting curve analysis and gel electrophoresis to identify non specific amplicons. Optimize your primer design to avoid such artifacts or use validated pre-designed primers.
PCR efficiency is <90%	PCR inhibitors present in a reaction mixture. Re-purify your template DNA. PCR conditions are suboptimal. Verify the primer concentrations. Verify storage conditions of qPCR master mix. Primer design. Verify your primer design, use primer design programs or validated pre-designed primers. Avoid designing primers in regions with high DNA secondary structure.

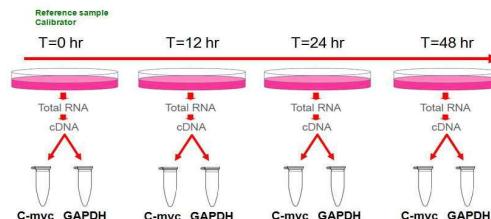


### Standard Sample

- When slope =  $-3.32$  Efficiency = 100%
- The optimal efficiency range: 90-110%
- R-value should be  $\geq 0.99$

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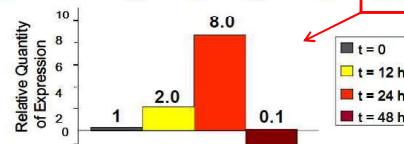
## Relative Quantitation (comparative Ct)



- ✓ Target gene: *c-myc*
- ✓ Reference gene
- (Endogenous control): **GAPDH**
- ✓ Calibrator : T= 0 hr

### $\Delta\Delta C_t$ Calculations (Comparative Ct)

	c-Myc	GAPDH	$\Delta C_t$	$\Delta\Delta C_t$
T=0 (calibrator)	25	10	15	0
T=12hr	24	10	14	-1
T=24hr	23	11	12	-3
T=48hr	28	10	18	3



#### step 1: Normalization to endogenous control

Sample: Ct *c-myc* - Ct *GAPDH* =  $\Delta C_t$  sample

Calibrator: Ct *c-myc* - Ct *GAPDH* =  $\Delta C_t$  calibrator

#### step 2: Normalization to calibrator sample

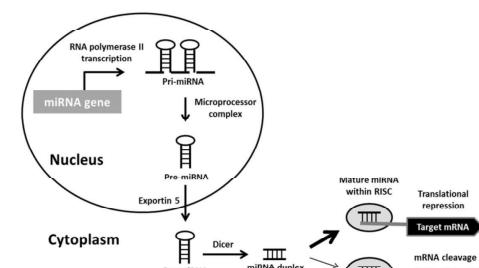
$\Delta C_t$  Sample -  $\Delta C_t$  Calibrator =  $\Delta\Delta C_t$

#### step 3: use the formula

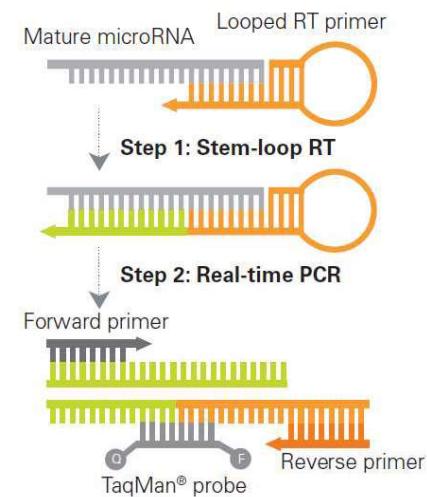
$$2^{-\Delta\Delta C_t}$$

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## TaqMan microRNA Assays



- Each assay contain:
- miRNA-specific RT primer
- miRNA-specific TaqMan Assay
- human, mouse, rat, *Drosophila*, *C. elegans*, and *Arabidopsis* etc.
- available; coverage for miRBase v.21



Front Cell Neurosci, Oct 2013 vol.7, article178  
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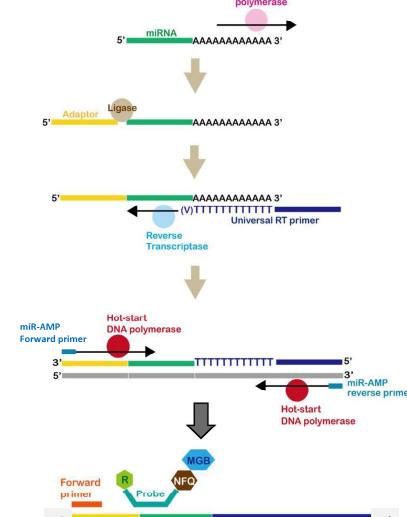
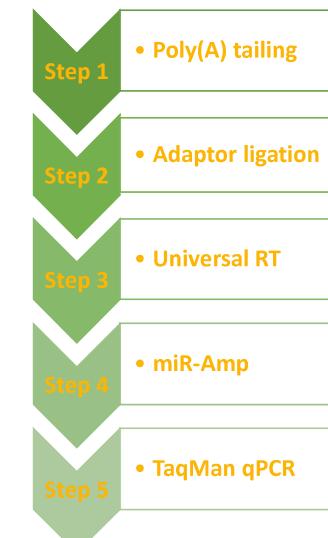
## TaqMan Advanced miRNA Assays

- superior sensitivity in tissue, serum, and plasma
- 1 ng of total RNA from tissue or 2  $\mu$ L of eluant from serum or plasma
- Have multiple miRNA targets from a single amplified sample
- TaqMan Advanced miRNA Assays is compatible with TaqMan advanced miRNA cDNA synthesis kit



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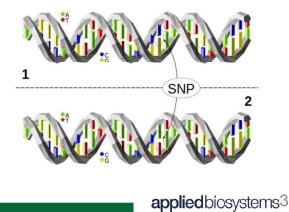
## TaqMan Advanced miRNA Assays: workflow



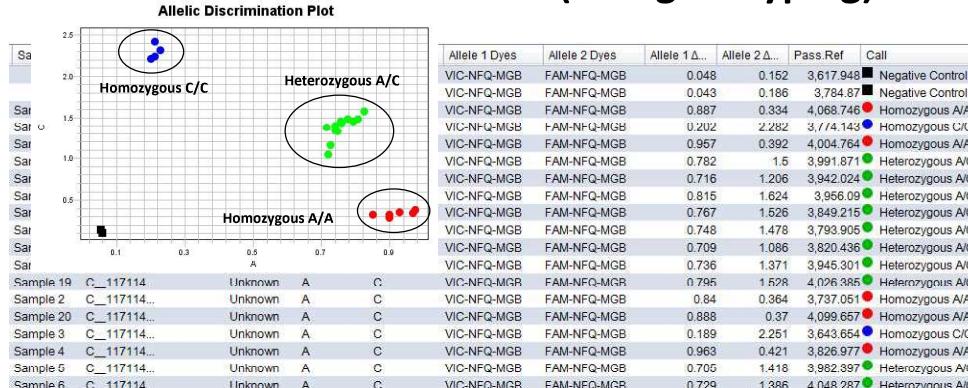
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## What are Single Nucleotide Polymorphism (SNP)?

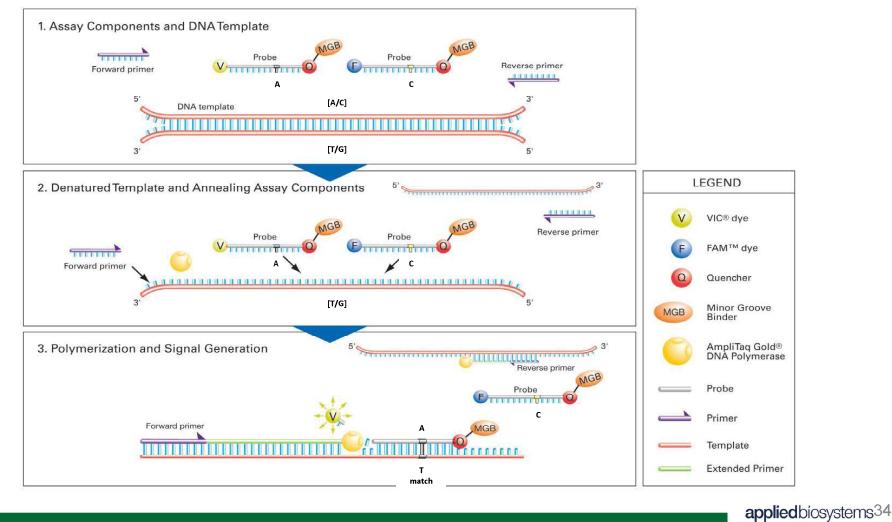
- Diploid organisms – 2 sets of chromosomes
- Each person receives 1 allele from each parent.
- If both alleles are the same, the person is **homozygous** for that gene.
- If the alleles differ, the person is **heterozygous** for that gene.
- They occur once in every 300 nucleotides on average.
- The rarer allele having a frequency of at least 1%.



## Allelic Discrimination Plot (SNP genotyping)



## TaqMan Probe System (Single Nucleotide Polymorphism)



## Application of TaqMan Assays and SYBR systems

TaqMan assays	SYBR Green
➤ Gene expression analysis (ex: Genetically Modified Organism, GMO)	
➤ MicroRNA and noncoding RNA analysis	➤ Gene expression analysis
➤ Drug metabolism genotyping ➤ SNP genotyping ➤ Somatic mutation detection	
➤ Pathogen Presence/ Absence	
➤ Protein expression	

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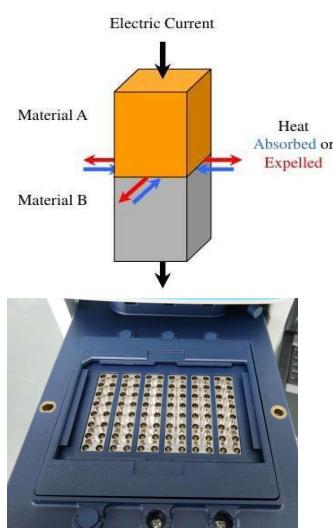
## The StepOnePlus Real-Time PCR System



## Design and Feature

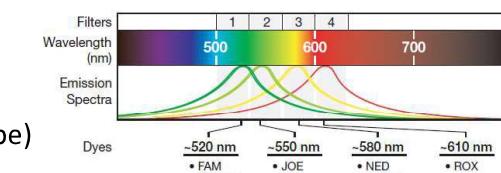


## Block



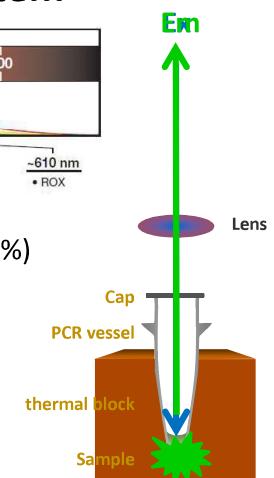
Tech.	→ Ramping	← Volume
1. Peltier effect	1. 鍍鉻合金基座 不同導體 電流改變 热能吸收 热電效應 精確控溫	1. 反應管 (0.1 mL) 96 well plate 8-tube strip single tube <b>*耗材需使用“Optical”</b>

## StepOne™ Plus Real-Time PCR System



### Common Features

- Fast and Standard ramping in the same block (1-100%)
- Single LED excitation
- Supported volumes are 10-30 µL
- Standalone or co-located
- Remote monitoring - LAN



## Supported Consumables

● for 0.1 mL Optical tube strip/single tube  
✓ 管子及蓋子上請勿標記任何記號  
避免墨色脫落沾到 PCR block 增加背景值

more than 48 reactions/run

✓ 96-well plate +  
Optical adhesive film

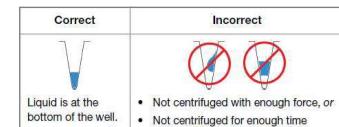
less than 48 reactions/run

✓ 使用八連排或單管必須將其置放在 MicroAmp™ 96-Well Tray  
✓ 對稱放置，避免加熱板上升時，受力不均，造成機械性的損傷。

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## 上機前/時之注意事項

- ✓ 光學封膜/瓶蓋是否平整緊密貼附，且表面無指紋或其他髒汙沾黏。
- ✓ 確認已將 reaction mixture 完全離心至反應管底部，且無氣泡產生 (Centrifuge the tubes for 2 min at less than 1500 rpm)
- ✓ 上機時，反應管放置位置應以“平行、偶數、對稱”為原則。
- ✓ 為了避免卡盤，不建議8連排剪裁後上機使用。

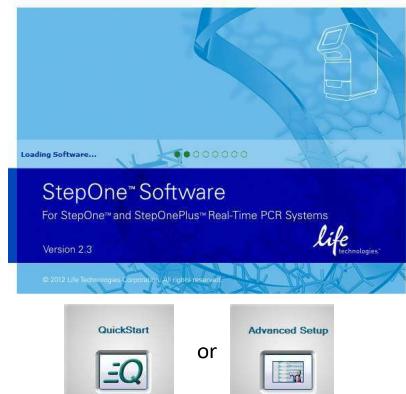


## SetUp

### 1. PC free



### 2. PC controlled



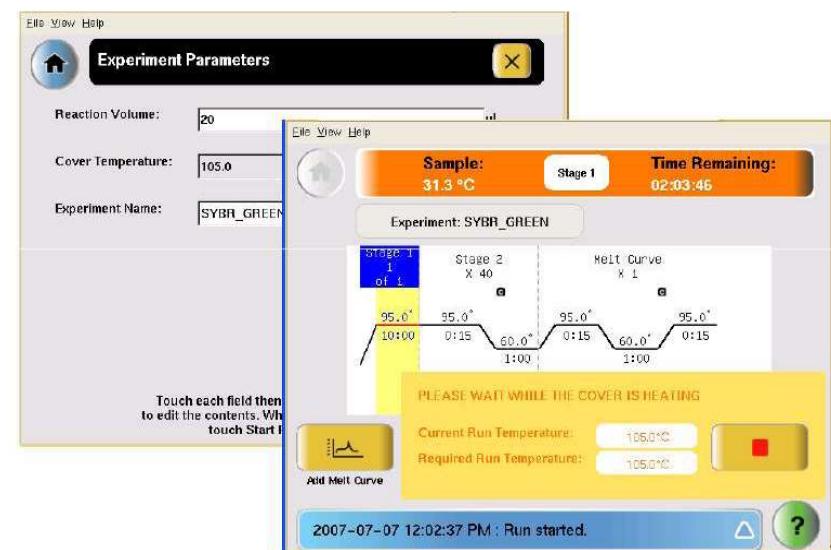
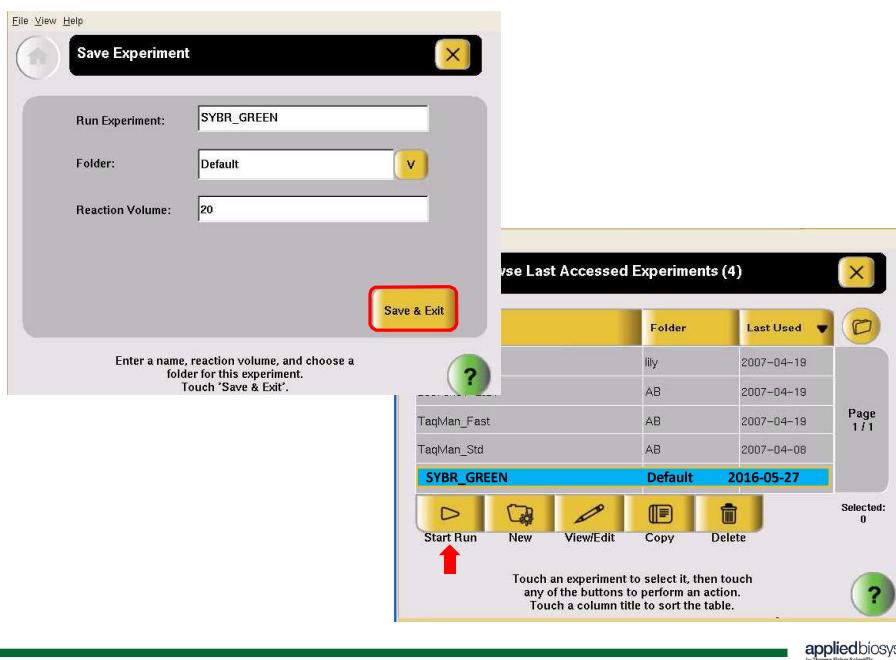
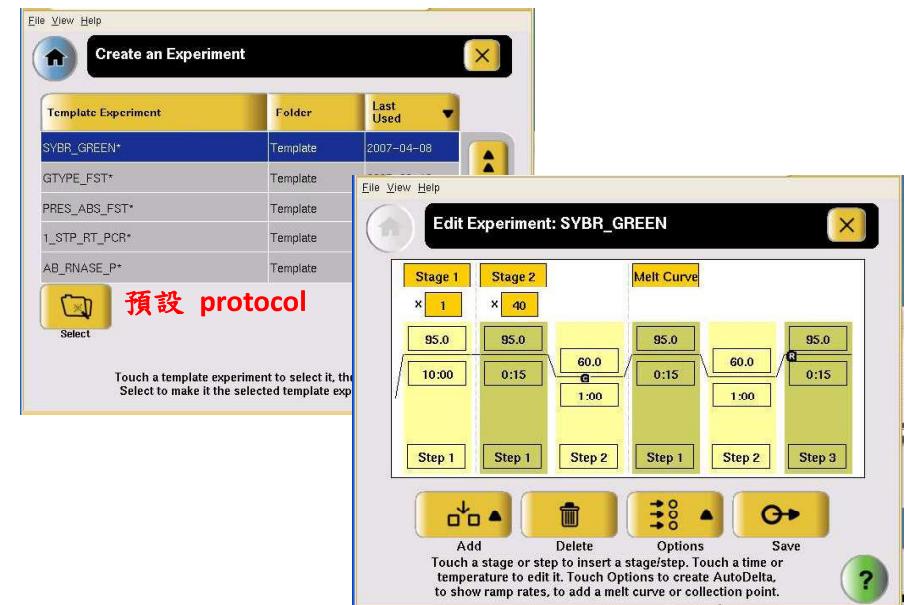
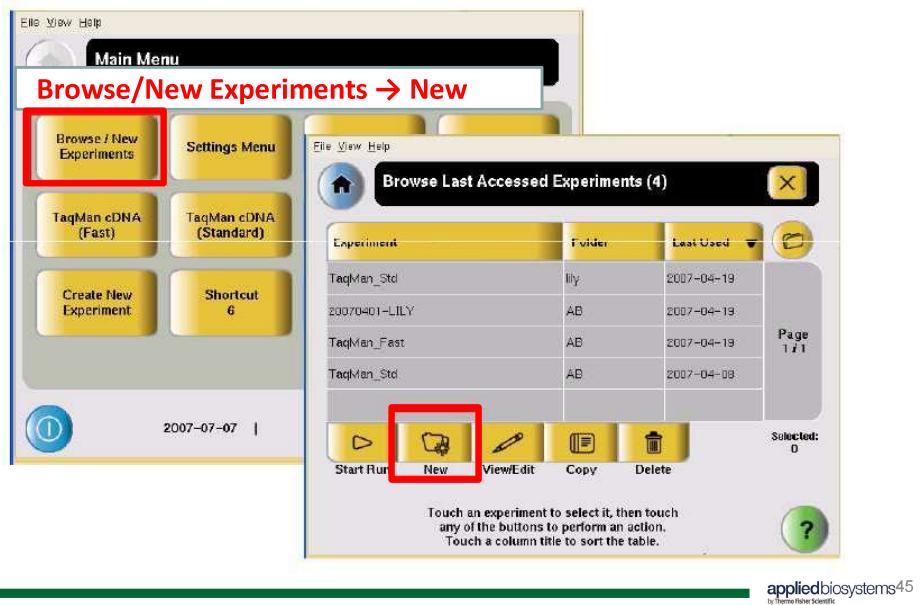
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## Standalone (PC-free)



1. Start the run from the touch screen.
2. Download the file (.eds) to your PC. (主機暫存最後一筆檔案)
3. Analyze your data.

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## StepOne Software v2.3 軟體操作



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## SetUp

### 1. PC free

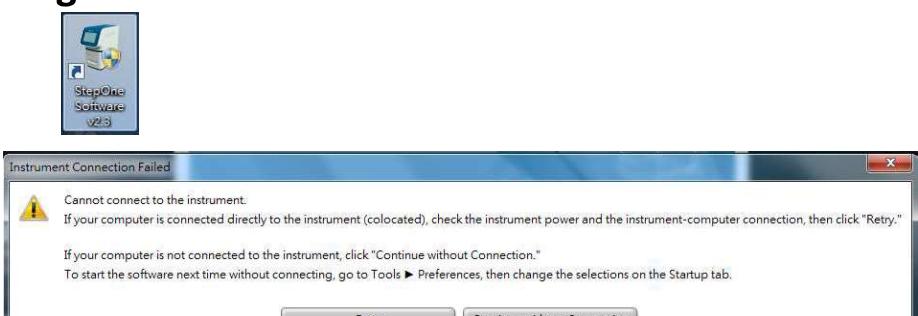


### 2. PC controlled



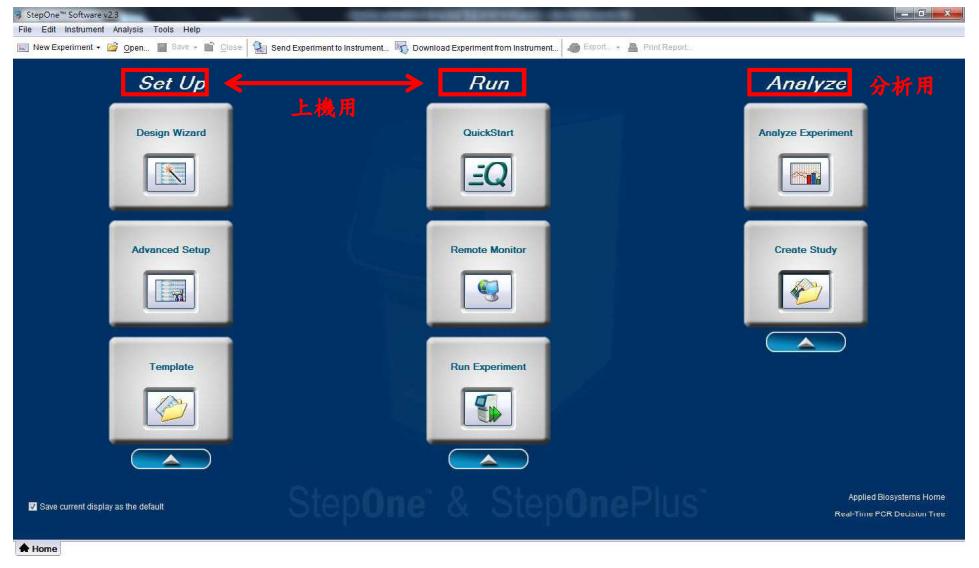
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## Login the software

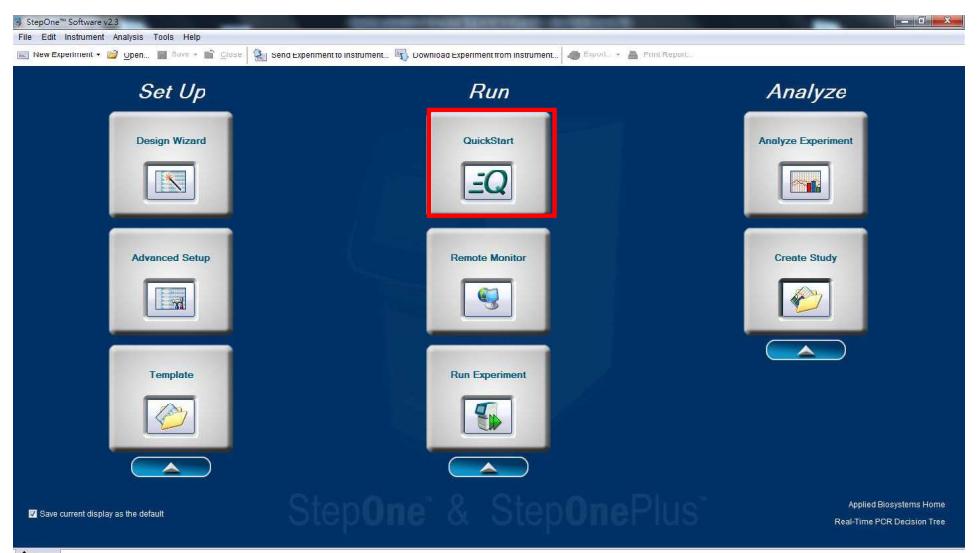


\* Analyze data -> Continue without Connection

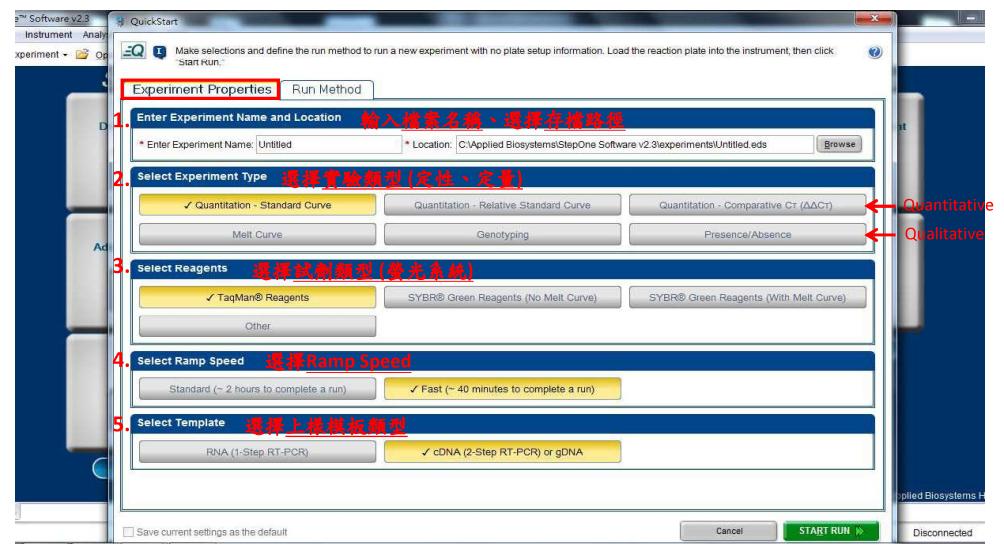
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## Run the experiment



### Run (QuickStart) : Experiment Properties



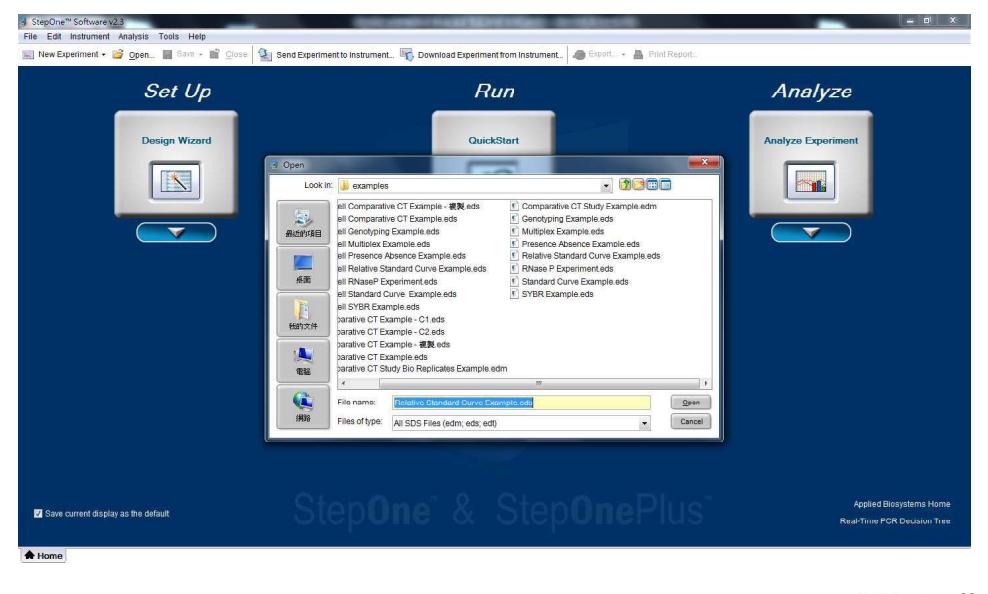
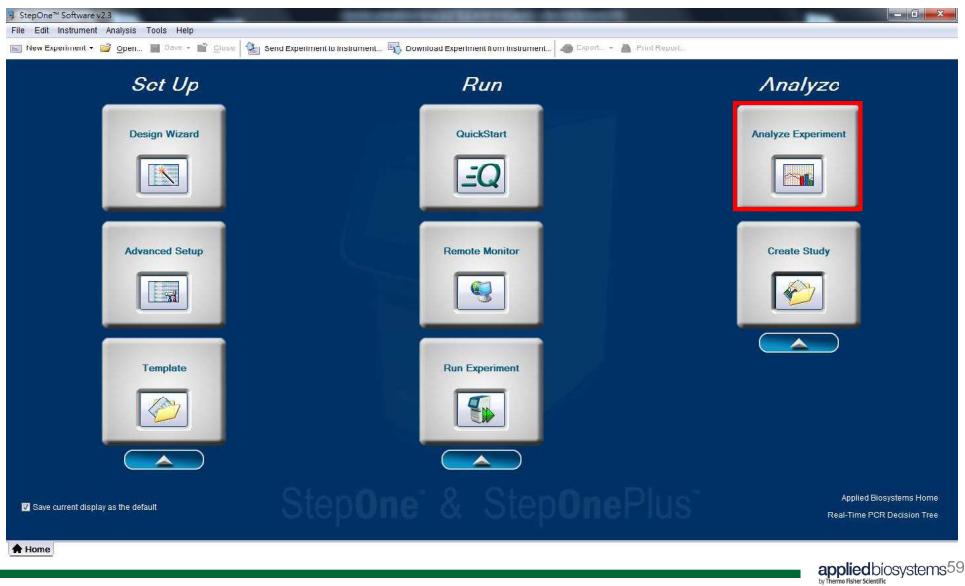
## Run (QuickStart) : Run Method



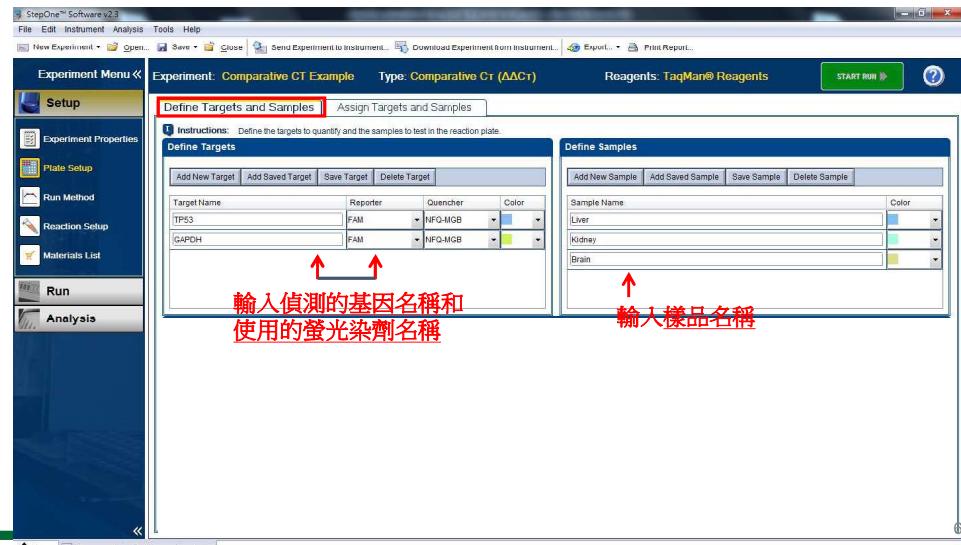
## Analyze the data



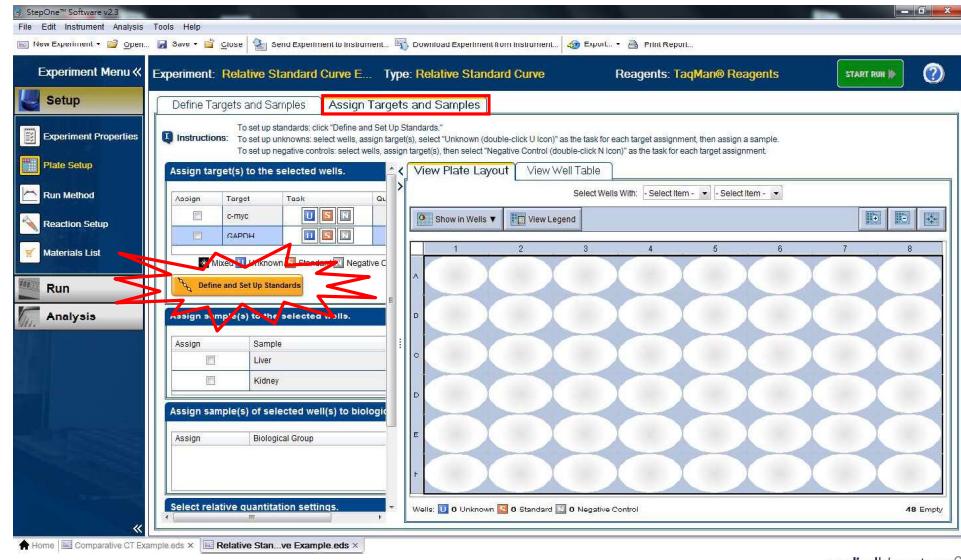
## Analyze: Analyze Experiment



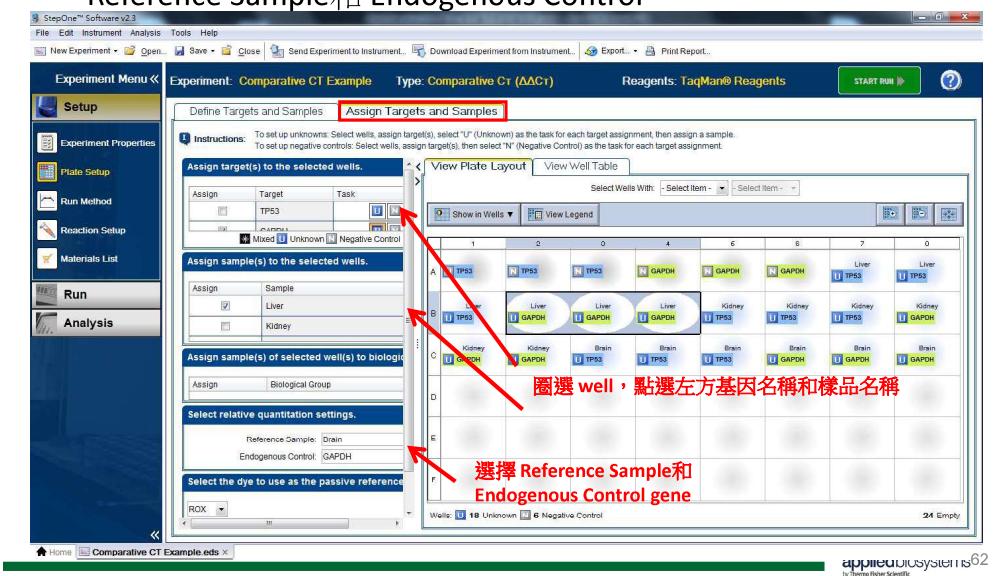
## Setup: plate Setup → Define Targets and Samples 定義偵測的基因和樣品名稱



## Setup: plate Setup → Assign Targets and Samples 定義標準品樣品位置、濃度



## Setup: plate Setup → Assign Targets and Samples 定義基因和樣品位置、設定 Reference Sample和 Endogenous Control



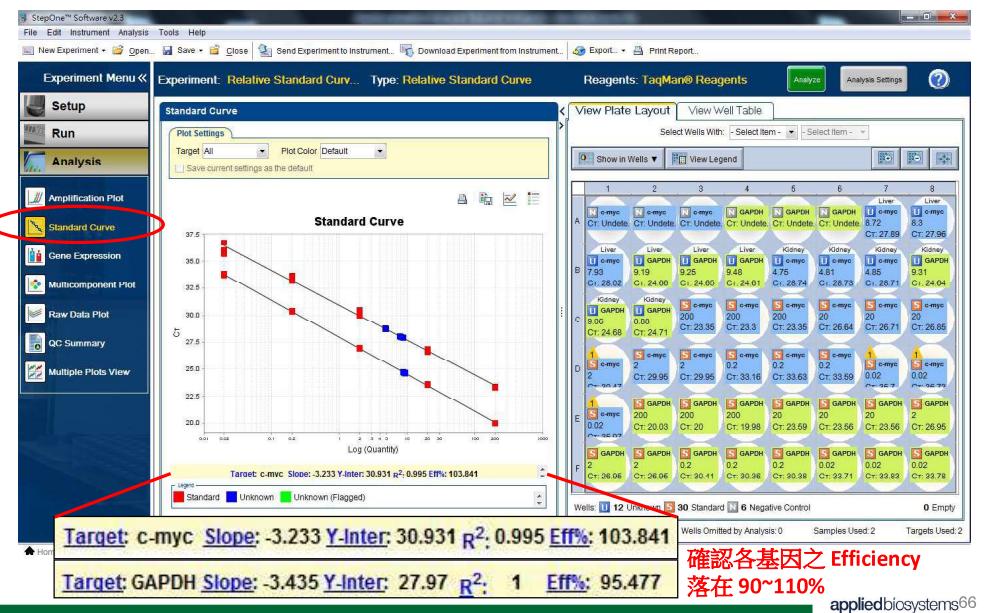
## Analysis: Melt curve (SYBR Green only)



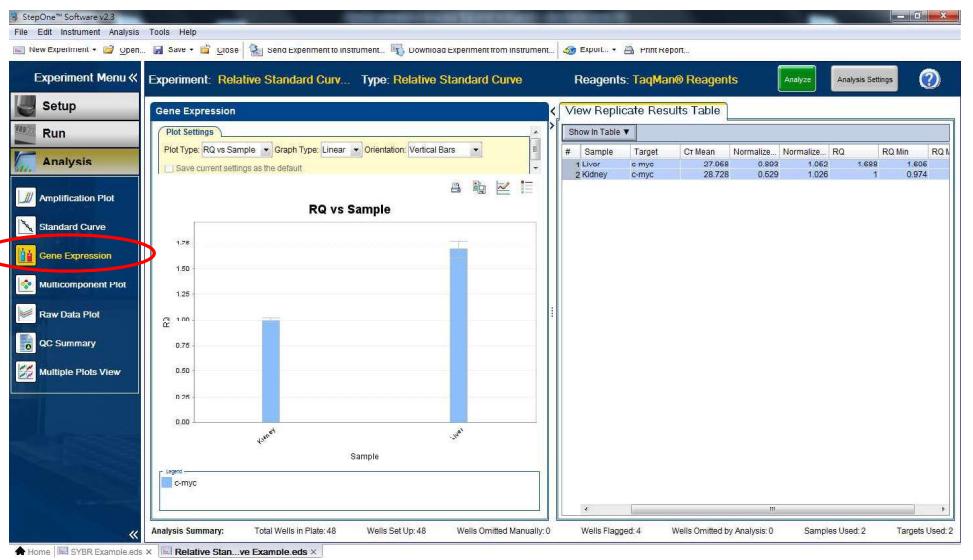
## Analysis: Amplification plot



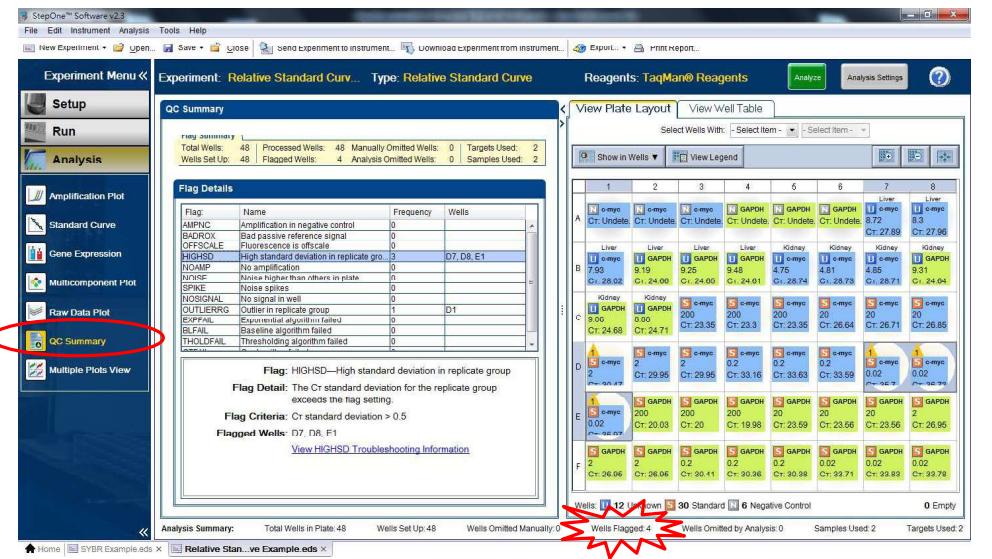
## Analysis: Standard curve



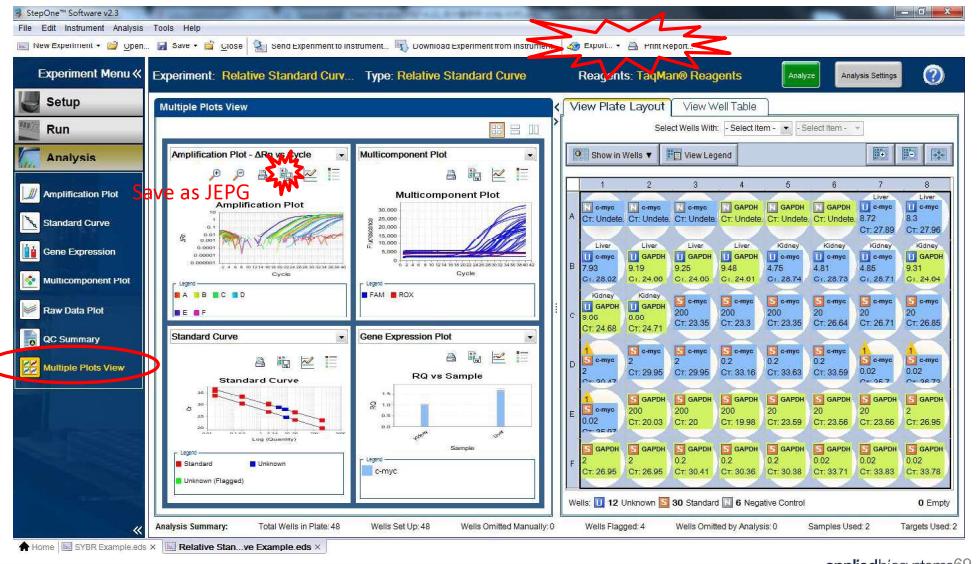
## Analysis: Gene Expression



## Analysis: QC Summary → 協助 troubleshooting



## Analysis: Multiple Plots View → Export to Excel or save as JPEG

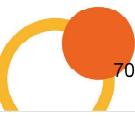
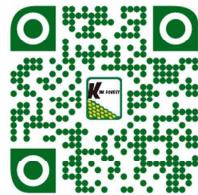


# Thank you for your attention!

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